RNAi & resistance against African SP weevils



Prof. Guy Smagghe

Research Group *Insect Physiology* & *Pest Control*

Dept. Crop Protection Fac. Bioscience Engineering

http://insects.ugent.be



African sweet potato weevil Cylas puncticollis & C. brunneus







- •Founded in <u>1817</u>. Now one of the leading institutions of higher education and research in *the Lower Countries* and in Europe. It distinguishes itself as a <u>socially committed and pluralistic university</u> in a broad international perspective.
- •Place <u>85 in Shanghai ranking</u> (Academic Ranking of 1000 World Universities); first of Belgium
- •Full university with <u>11 faculties</u>, 130 departments and <u>38,000 students</u>.
- 5 <u>Doctoral Schools</u> for doctoral researchers.

1. RNAi mechanism2. Project outline with WPs

3. First results







The Nobel Prize in Physiology or Medicine 2006 – Andrew Z. Fire, Craig C. Mello





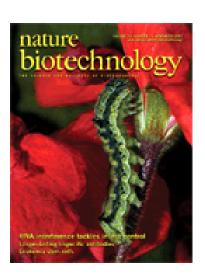
NATURE BIOTECHNOLOGY VOLUME 25 NUMBER 11 NOVEMBER 2007

Control of coleopteran insect pests through RNA interference

James A Baum¹, Thierry Bogaert², William Clinton¹, Gregory R Heck¹, Pascale Feldmann², Oliver Ilagan¹, Scott Johnson¹, Geert Plaetinck², Tichafa Munyikwa¹, Michael Pleau¹, Ty Vaughn¹ & James Roberts^{1,3}

Silencing a cotton bollworm P450 monooxygenase gene by plant-mediated RNAi impairs larval tolerance of gossypol

Ying-Bo Mao^{1,2}, Wen-Juan Cai^{1,2}, Jia-Wei Wang^{1,2}, Gao-Jie Hong^{1,2}, Xiao-Yuan Tao^{1,2}, Ling-Jian Wang¹, Yong-Ping Huang¹ & Xiao-Ya Chen¹



NATURE BIOTECHNOLOGY VOLUME 25 NUMBER 11 NOVEMBER 2007

Control of coleopteran insect pests through RNA interference

James A Baum¹, Thierry Bogaert², William Clinton¹, Gregory R Heck¹, Pascale Feldmann², Oliver Ilagan¹, Scott Johnson¹, Geert Plaetinck², Tichafa Munyikwa¹, Michael Pleau¹, Ty Vaughn¹ & James Roberts^{1,3}









Western corn rootworm, Diabrotica virgifera virgifera



Reduction in root damage



Contents lists available at ScienceDirect

Journal of Insect Physiology

journal homepage: www.elsevier.com/locate/jinsphys



Review

Mechanisms of dsRNA uptake in insects and potential of RNAi for pest control: A review

Hanneke Huvenne, Guy Smagghe*

Laboratory of Agrozoology, Department of Crop Protection, Ghent University, Coupure Links 653, 9000 Ghent, Belgium



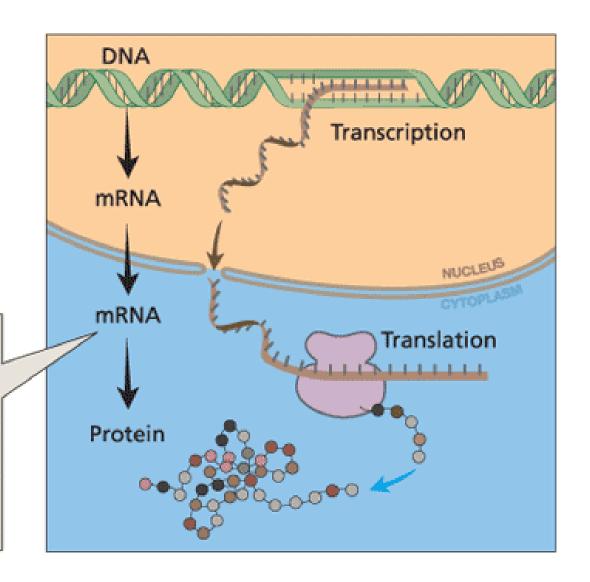
Central dogma

The genetic information in doublestranded DNA is transcribed into single-stranded messenger RNA (mRNA) in the cell nucleus and subsequently translated into protein in the cytoplasm.



RNA interference

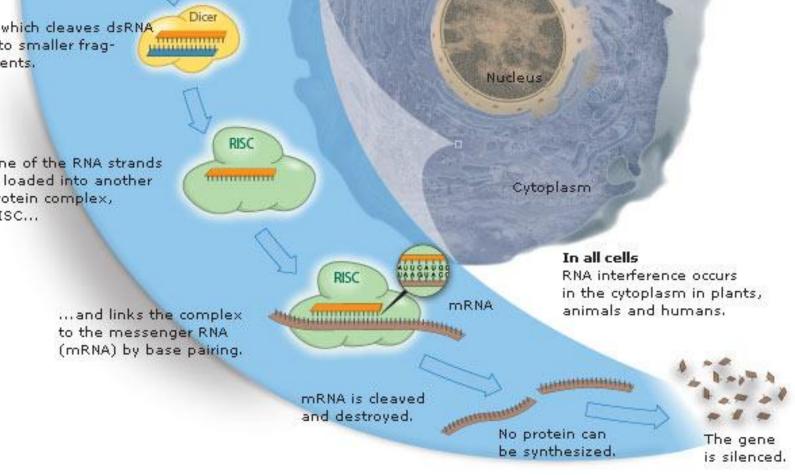
The mRNA is destroyed before it is translated into protein.



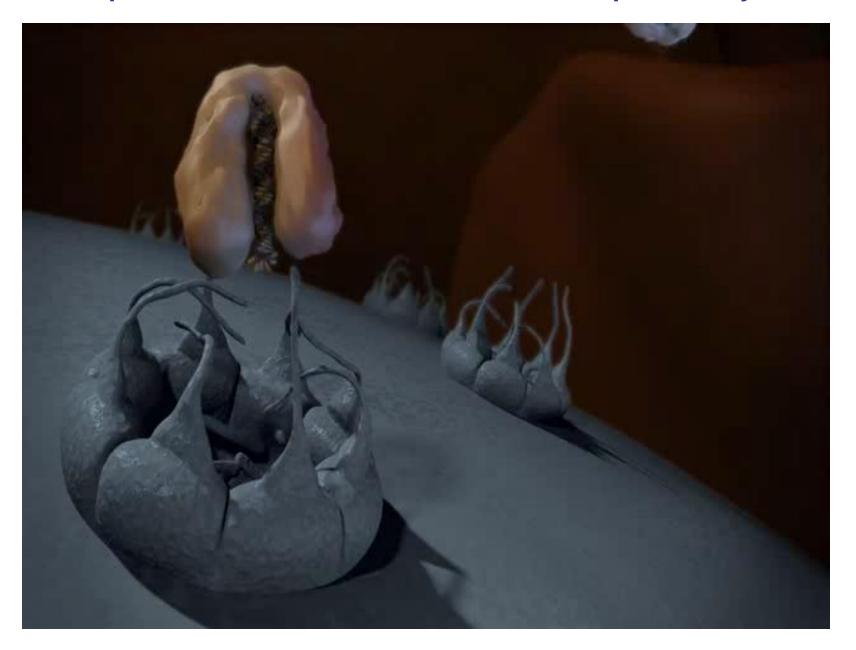
<u>RNAi</u>

- Fundamental research
- -> functional genomics
- •Applications:
- ->insect pest control
- ->protection of beneficial insects (e.g. bees) against pathogenic viruses

Tailor-made ds/siRNA molecules RNA interference, RNAi Double-stranded RNA triggers gene silencing. -> degrade specific mRNA Double-stranded RNA (dsRNA) binds to a protein complex, Dicer... ARREST MARKET THE PROPERTY OF THE PERSON NAMED IN COLUMN TWO PARTY OF THE PERSON NAMED IN COLUMN TWO PARTY OF THE PERSON NAMED IN COLUMN TO THE PERSON NAMED dsRNA Dicer ...which cleaves dsRNA into smaller fragments. Nucleus RISC One of the RNA strands Terrorion to the Contract of t is loaded into another Cytoplasm protein complex, RISC ...



RNAi: a process where dsRNA silences the complementary mRNA



1. RNAi mechanism2. Project outline with WPs

3. First results





PROJECT DETAILS:

- North-South project
- Collaboration CIP, UGent (Prof. Guy Smagghe, Prof. Godelieve Gheysen, Dr. Marc Ghislain)
- •4 years; started March 2013
- •4 WPs:
 - •WP1-Biology and maintenance of colony of African SP weevils Cylas puncticollis and C. brunneus in the laboratory
 - •WP2-Trancriptome analysis and selection of best candidate genes in Cylas puncticollis and C. brunneus for RNAi
 - •WP3-Transformation of African cultivars of sweetpotato with best candidate(s) of insect-specific dsRNA
 - •WP4-Testing the efficacy of transformed African sweetpotato cultivars to resist against an infection of African SP weevils of Cylas sp.



1. RNAi mechanism2. Project outline with WPs

3. First results







We started

- 2+2 yrs sandwich PhD grant North-South by UGent
- Collaboration with CIP in Uganda/Kenia, and Venganza



Katterinne Prentice

WP1-Biology and maintenance of colony of African SP weevils Cylas puncticollis and C. brunneus in the laboratory

- •Start with weevils from NaCRRI lab in Uganda
- Colony at Ghent University on fresh sweet potatoes. Controlled incubators Sanyo at 25°C 75% RH, 16L:8D
- Also attempts to keep colony on artificial diet

WP2-Trancriptome analysis and selection of best candidate genes in Cylas puncticollis and C. brunneus for RNAi

- -Sequencing at Genomic Service of North Caroline State Univ., USA
- -Gene annotation at NCSU and UGent
- -Selection of c.20 essential target genes (+ housekeeping genes for qPCR)

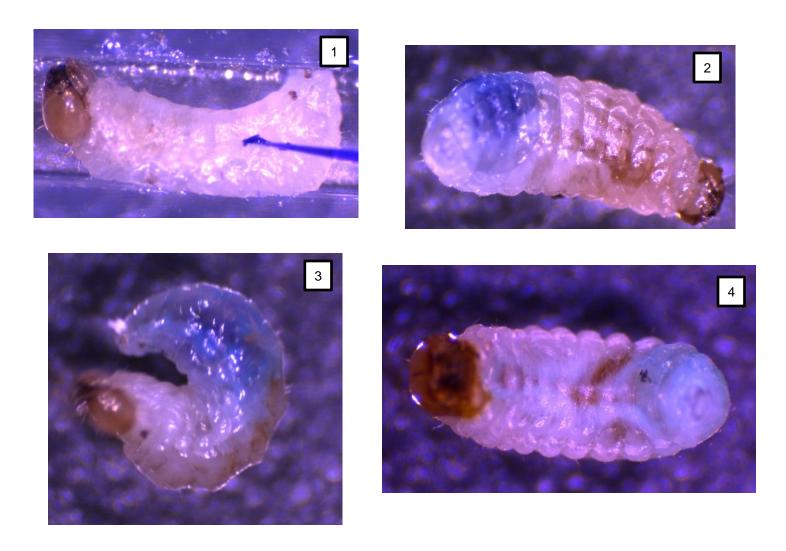
	Gene	C. p.	C. b.
1	Vha68-2 F1 ATP synthase beta subunit	X	X
2	V/A-type ATP synthase catalytic subunit A	X	X
3	Synaptobrevin, isoform A	X	X
4	Pfk phosphofrutokinase		X
5	adenylate kinase-2	X	X
6	Focal adhesion kinase isoform D		X
7	gamma-coatomer protein, isoform C	X	
8	delta-coatomer protein, isoform A	X	
9	alpha-coatomer protein, isoform D	X	
10	TBP-associated factor 1, isoform D	X	X
11	lethal (2) NC136, isoform B	X	X
12	Proteasome 20 kD subunit	X	X
13	DNA polymerase interactin tpr containing protein of 47 Kd, isoform B	X	X
14	alpha-Adaptin, isoform A	X	X
15	Mad1	X	X
16	Ubiquitin conjugating enzyem E2	X	
17	RNA pol beta subunit	X	X
18	RNA helicase	X	X
19	ribosomal protein S13e	X	X
20	DNA polymerase alpha 50 kD		X
21	vATPase A		X
22	vATPase D	X	X
23	RPL19		X
24	Snf7		

WP2-Trancriptome analysis and selection of best candidate genes in Cylas puncticollis and C. brunneus for RNAi

- -Sequencing at Genomic Service of North Caroline State Univ., USA
- -Annotation at NCSU and UGent
- -Selection of c.20 essential target genes (+ housekeeping genes for qPCR)
- -Synthesis of dsRNAs for these c.20 target genes
- -Screening insecticide activity of c.20 dsRNAs in insect bioassays by nano-injection

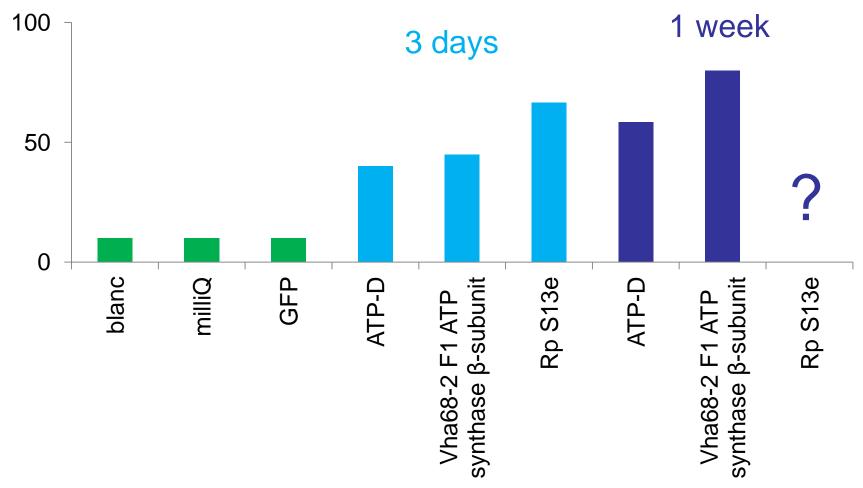


Cylas puncticollis, early 3rd larval instar



(1) Injection with blue dye. (2) after 0-1 min. (3) 1 h (4) 1 day.

First data with % insect mortality by nano-injection



- dose dsRNA injected = 100 ng/mg
- •total N=190 third instars C.p.
- no info on transcription silencing yet

Blanc water control



dsRNA Rp S13e



WP2-Trancriptome analysis and selection of best candidate genes in Cylas puncticollis and C. brunneus for RNAi

- -Sequencing at Genomic Service of North Caroline State Univ., USA
- -Gene annotation at NCSU and UGent
- -Selection of c.20 essential target genes (+ housekeeping genes for qPCR)
- -Synthesis of dsRNAs for these c.20 target genes
- -Screening insecticide activity of c.20 dsRNAs in insect bioassays by nano-injection
- -Phenotypes are focusing on lethal effects, larval development, growth and reproduction
- -Analyze efficacy of transcription silencing by qPCR
- -Selection of c.5 best candidates
- -Testing insecticide potency of c.5 best dsRNAs by feeding
- -Testing insecticidal potency of combinations
- -Testing insecticide potency of c.5 best dsRNAs with insects collected from field

WP3-Transformation of African cultivars of sweetpotato with best candidate(s) of insect-specific dsRNA

->later in 2014



Thank you!

 Many enthusiastic MSc (87) and PhD students (18 finished and 22 ongoing)

Funding:

- Belgian Fund for Scientific Research (FWO)
- Special Research Fund of Ghent University

