



Germplasm Virus Indexing: Supporting ISO/IEC 17025 Accreditation

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April 12, 2012



Outline

- **Introduction**
- **ISO/IEC 17025 accreditation**
- **Virus indexing procedure**
- **Analysis of indexing results**
- **Research derived from indexing**
- **Maintaining ISO accreditation**
- **New molecular tests (validation)**



Outline

➤ Introduction



Fact:

CIP is the custodian of world's largest *in vitro* germplasm



Number of accessions maintained in CIP's *in vitro* germplasm (April 04, 2012)

Sweetpotato: 5,757

Potato: 8,557

Data provided by B. Zea



Number of accessions distributed internationally

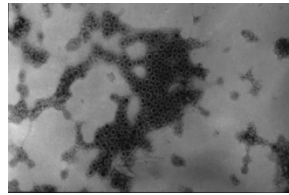
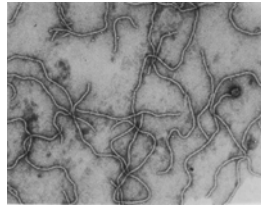
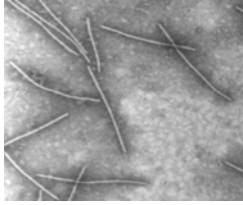
| Crop | Year | | | | | | Average |
|--------------|------|-------|-------|-------|------|-------|---------------|
| | 2006 | 2007 | 2008 | 2009 | 2010 | 2011 | |
| Sweetpotato | 291 | 939 | 394 | 632 | 206 | 899 | 560 |
| Potato | 413 | 172 | 999 | 512 | 723 | 611 | 571 |
| TOTAL | 704 | 1,096 | 1,341 | 1,122 | 886 | 1,510 | 1, 131 |

Data provided by R. Vollmer, E. Grande and E. Rojas

Virus-free *in vitro* germplasm (HS2)



Viruses affecting sweetpotato



> 30 viruses





Symptoms induced by virus infection





Symptoms induced by virus infection





Symptomless infection





Virus Indexing

- It combines the knowledge of the viruses with the methodologies available for their detection (to ensure effective safe movement of germplasm)



Virus Indexing

- It combines the knowledge of the viruses with the methodologies available for their detection (to ensure effective safe movement of germplasm)
- Guidelines from the “Safe Movement of Sweet Potato Germplasm” (1989)



It can be revised and modified over time



Virus Indexing

- It combines the knowledge of the viruses with the methodologies available for their detection (to ensure effective safe movement of germplasm)
- Guidelines from the “Safe Movement of Sweet Potato Germplasm” (1989)
- Guideline on our SP virus indexing includes:
 - Observation of symptoms on SP plants
 - Grafting to *Ipomoea setosa*
 - Serological test (NCM-ELISA) from grafted plants
 - Molecular test (PCR)



Outline

- Introduction
- **ISO/IEC 17025 accreditation**



Facts about CIP's ISO Accreditation:

- **CIP awarded the ISO/IEC 17025 accreditation on February 2008 and applies this ISO Standard for the management and distribution of CIPs germplasm.**

ISO/IEC 17025



Testing laboratory
No. 4229

**For safe and
secure germplasm
movement.**

Source: CIP's ISO Accreditation Website



Facts about CIP's ISO Accreditation:

- **CIP awarded the ISO/IEC 17025 accreditation on February 2008 and applies this ISO Standard for the management and distribution of CIPs germplasm.**
- **ISO/IEC 17025 is a worldwide Quality Standard that sets down requirements for the competence of testing laboratory.**

Management Requirements

- Complaints
- Control of records
- Corrective and preventive actions
- Document control
- General Requirements
- Improvement
- Internal audits
- Management review
- Non-conforming Work
- Purchasing of services and Supplies
- Review of requests for germplasm material
- Service to clients
- Subcontracting
- Technical Management and Quality Manager

Technical Requirements

- Accommodation and environment
- Copy of Quality Control procedures
- Germplasm Health Statement
- Handling of plant materials
- Measurement traceability
- Personnel and training
- Quality Control procedures
- Selection and validation of diagnostic methodology

Source: CIP's ISO Accreditation Website

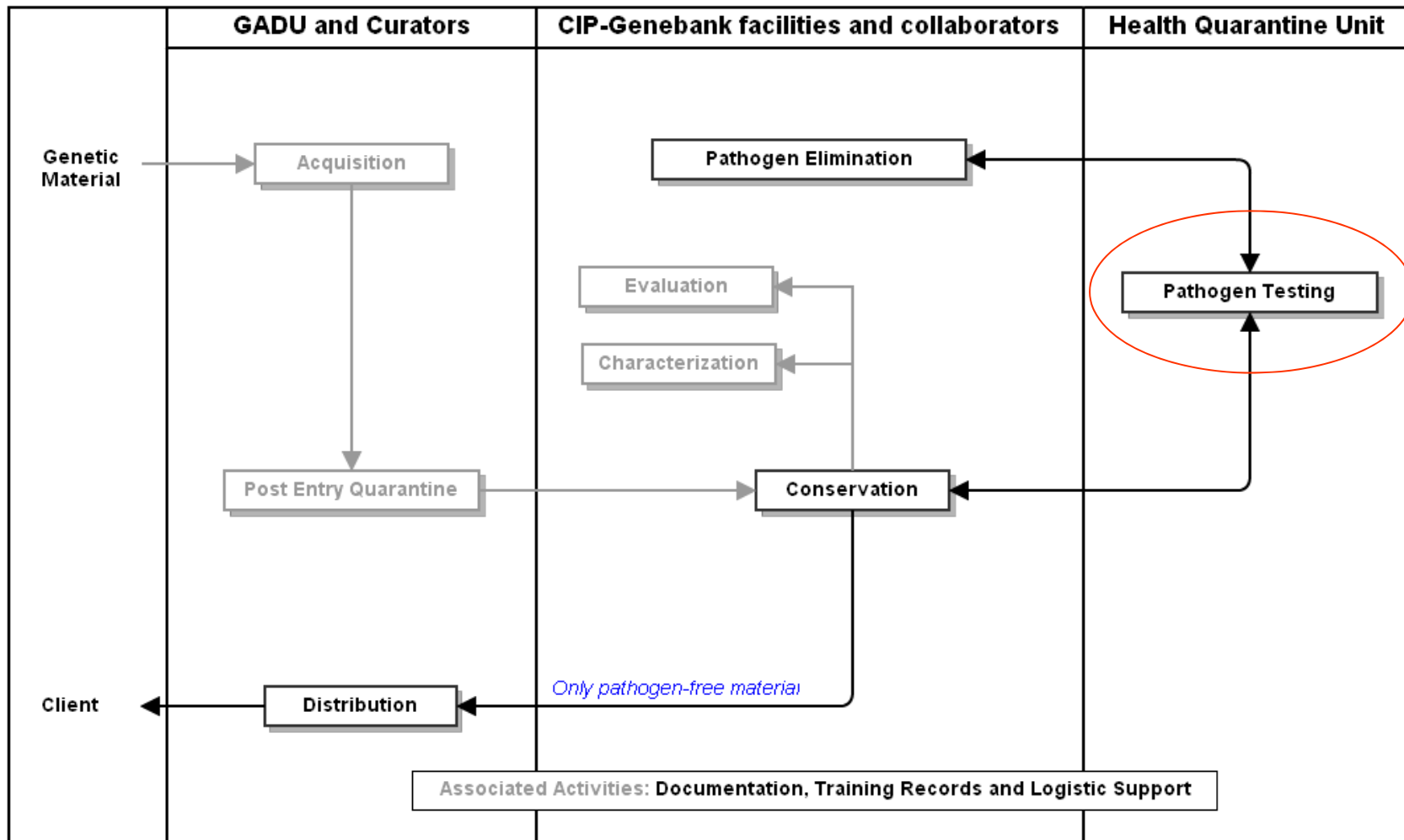


Facts about CIP's ISO Accreditation:

- **ISO/IEC 17025 is a worldwide Quality Standard that sets down requirements for the technical competency of testing laboratory.**
- **CIP awarded the ISO accreditation on 22 February 2008 and applies this ISO Standard the for management and distribution of CIP-HQ's germplasm.**
- **With ISO accreditation, CIP gives the users of the Genebank a visible assurance of the quality of the germplasm being distributed.**



WORKFLOW 1 (WF1) : Acquisition, Post Entry Quarantine, Consevation, Pathogen Testing, Pathogen Elimination and Distribution of materials in CIP-Genebank: Overview



Source: CIP's ISO Accreditation Website

In gray color: non-accredited process



List of Pathogen Detection In-Vivo Diagnostic Methodology

1. Test Covered by the UKAS Scope of Accreditation

| Methodology | Pathogens Detected |
|--------------------|--|
| <u>Potato</u> | |
| DAS-ELISA | PVX, PVY, PLRV, PVS, APMoV, APLV, PYV, AVB-O |
| NASH | PVT, PSTVd |
| <u>Sweetpotato</u> | |
| NCM-ELISA | SPFMV, SPLV, SPVG, SPMSV, SPMMV, SPCFV, C-8 virus, SPCSV, SPCV (former as SPCaLV), CMV |

2. Other Tests (to be incorporated into the Accreditation scope with time)

| Methodology | Pathogens Detected |
|--------------------|--|
| <u>Potato</u> | |
| PCR | Phytoplasmas, Begomoviruses transmitted by whiteflies, PVV |
| <u>Sweetpotato</u> | |
| PCR | Begomoviruses including SPLCV, Phytoplasmas |



Outline

- Introduction
- ISO/IEC 17025
- **Virus indexing procedure**



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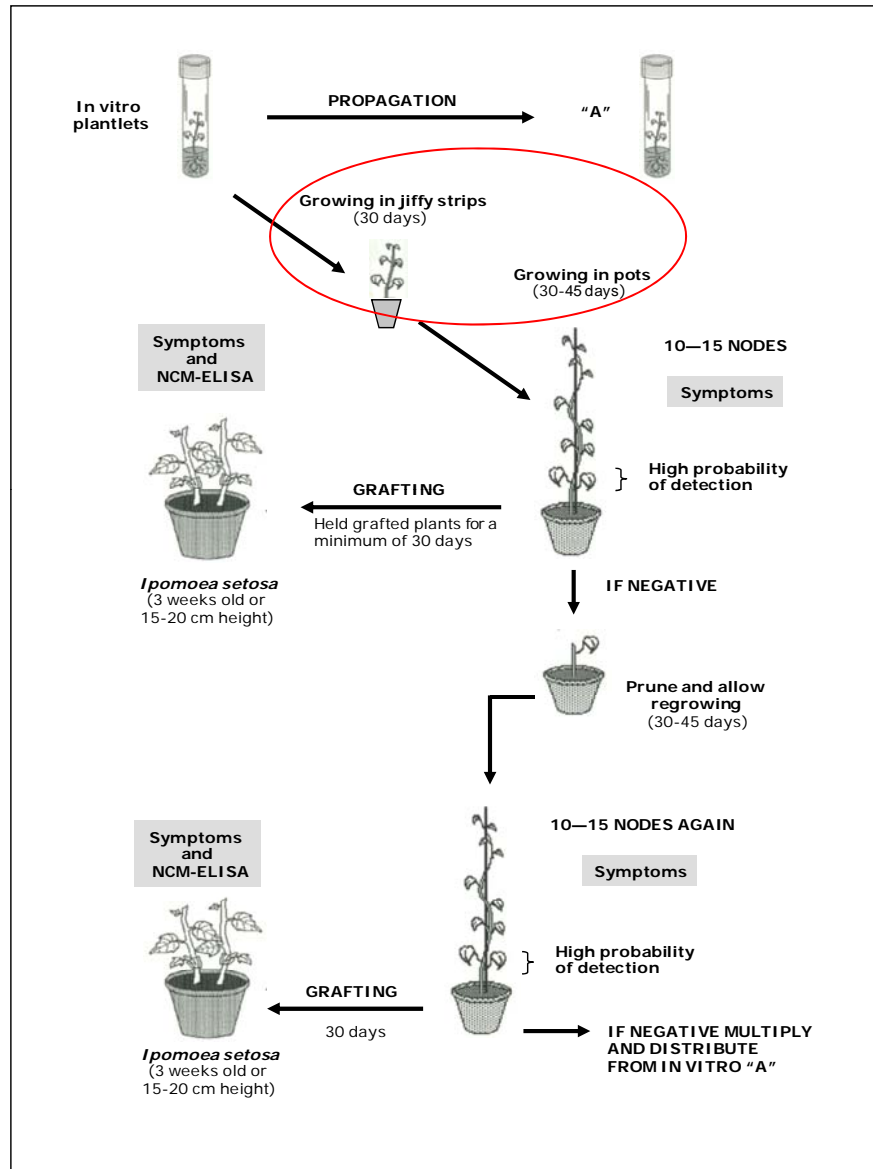
graph TD
    A[In vitro plantlets] -- PROPAGATION --> B((A))
    B -- "Growing in jiffy strips (30 days)" --> C[Growing in pots (30-45 days)]
    C -- "Symptoms" --> D[PCR is performed to detect begomoviruses (since June 2009)]
    C -- "10-15 NODES" --> E[High probability of detection]
    C -- "IF NEGATIVE" --> F[Prune and allow regrowing (30-45 days)]
    F -- "Symptoms" --> G[10-15 NODES AGAIN]
    G -- "High probability of detection" --> H[High probability of detection]
    G -- "IF NEGATIVE MULTIPLY AND DISTRIBUTE FROM IN VITRO A" --> I(( ))
    C -- "GRAFTING" --> J[Ipomoea setosa (3 weeks old or 15-20 cm height)]
    J -- "Held grafted plants for a minimum of 30 days" --> K[Symptoms and NCM-ELISA]
    G -- "GRAFTING" --> L[Ipomoea setosa (3 weeks old or 15-20 cm height)]
    L -- "30 days" --> M[Symptoms and NCM-ELISA]
  
```

The flowchart illustrates the indexing protocol for *Ipomoea setosa*. It begins with 'In vitro plantlets' which undergo 'PROPAGATION' to become 'A' (circled in red). 'A' is then grown in 'jiffy strips (30 days)' and then 'in pots (30-45 days)'. At this stage, 'Symptoms' are checked, and 'PCR is performed to detect begomoviruses (since June 2009)'. If 'NEGATIVE', the plant is 'Pruned and allowed regrowing (30-45 days)', then checked for 'Symptoms' again. If still 'NEGATIVE', it is 'Multiplied and distributed from in vitro A'. If 'POSITIVE' at either check, the plant is grafted onto 'Ipomoea setosa (3 weeks old or 15-20 cm height)' and held for '30 days' before being checked for 'Symptoms and NCM-ELISA'.

20



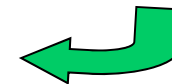
Sweetpotato virus indexing procedure



Indexing procedure for sweet potato viruses. NCM-ELISA is performed for 10 viruses (SPFMV, SPLV, SPVG, SPMSV, SPMMV, SPCSV, SPCFV, C-6, SPCV, and CMV). PCR is performed to detect begomoviruses.

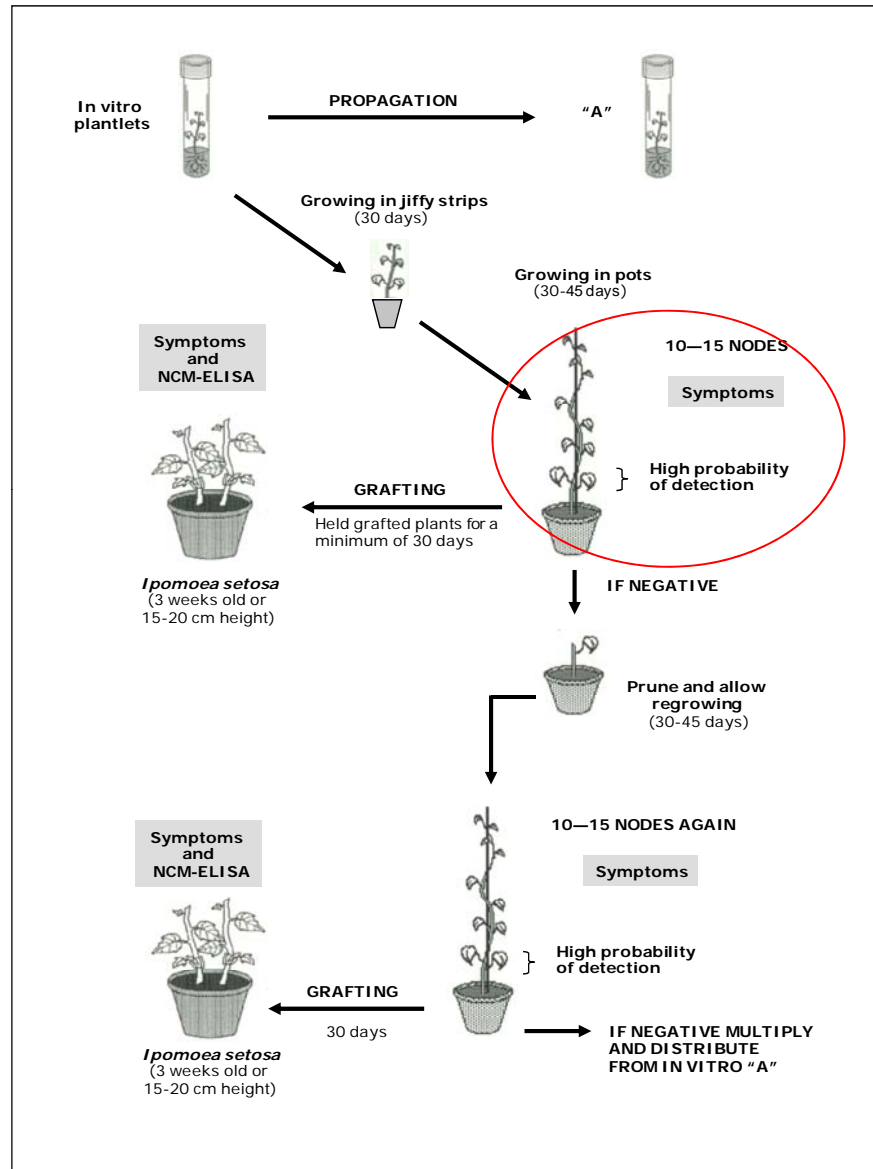


Growing plantlets in a greenhouse





Sweetpotato virus indexing procedure



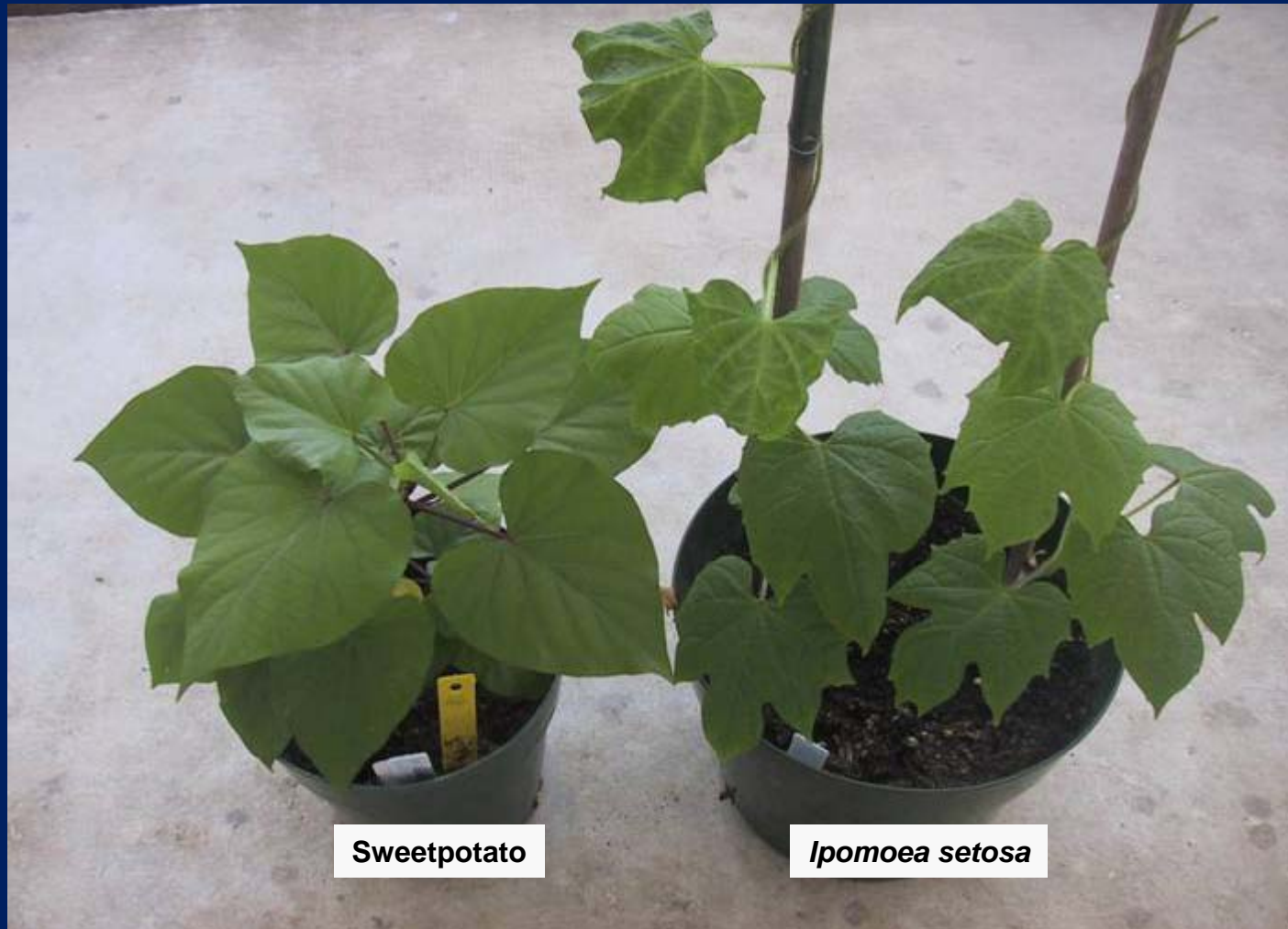
Indexing procedure for sweet potato viruses. NCM-ELISA is performed for 10 viruses (SPFMV, SPLV, SPVG, SPMSV, SPMMV, SPCSV, SPCFV, C-6, SPCV, and CMV). PCR is performed to detect begomoviruses.



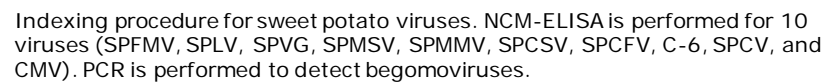
Symptoms observed in grown sweetpotato plants



Symptomless infection



- Low virus concentration in sweetpotato tissues (virus is not detected by serology from the SP plant)





Transmission of plant viruses

I. Mechanical and by contact

II. **Grafting** ← (universal method for transmitting viruses) — **Sweetpotato**

III. Vectors

Insects (aphids, whiteflies)

IV. Pollen and sexual seed



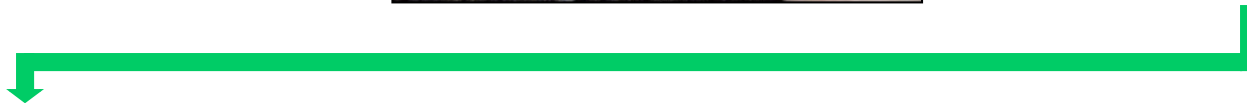
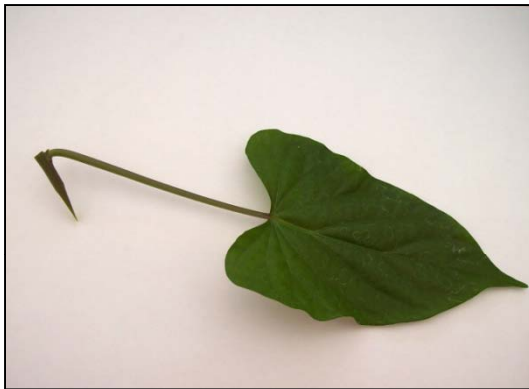
Ipomoea setosa



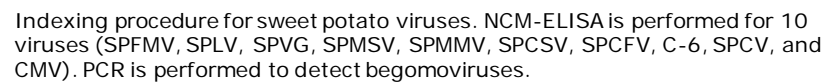
Universal indicator plant for viruses that infect sweetpotato



Wedge-grafting to *Ipomoea setosa*



Grafting: universal method for virus transmission





Symptoms recording and NCM-ELISA test



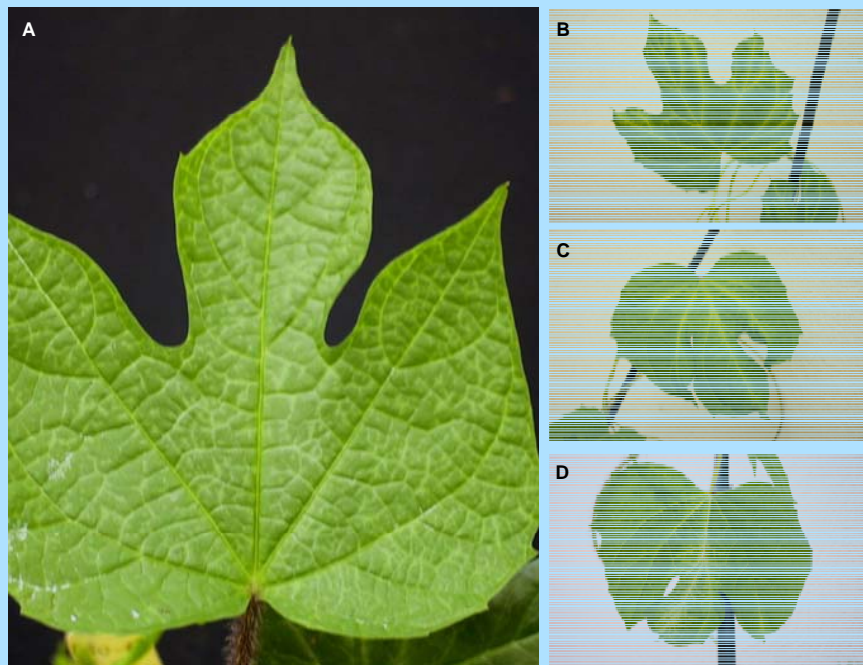
Preventive spraying of insecticides (controlling of whiteflies and aphids)



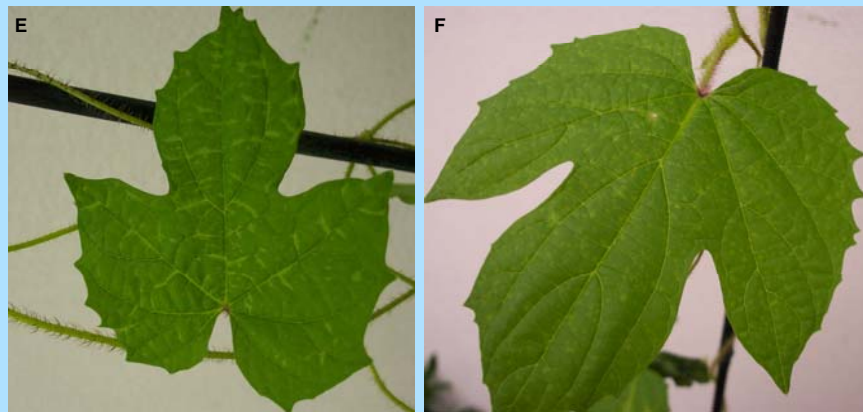
Symptoms1: Vein clearing

(Potyvirus)

SPFMV



SPVG

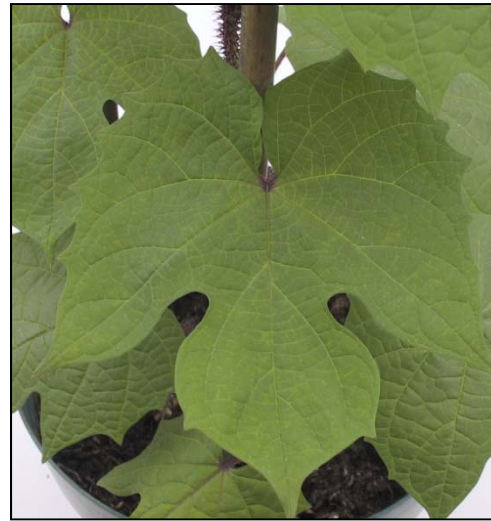


Unknown
(Negative in NCM-ELISA)



Symptoms 2: Chlorotic points

(Carlavirus)





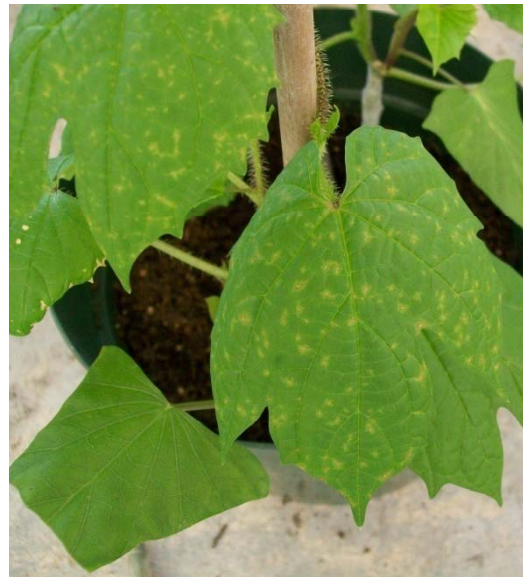
Symptoms 3: Vein clearing / rugosity





Symptoms 4: Vein necrosis/ necrotic points or spots

(Cavemovirus/ Solendovirus)





Symptoms 5: Chlorosis/ interveinal chlorosis/ roll up/ roll down

(Begomovirus)

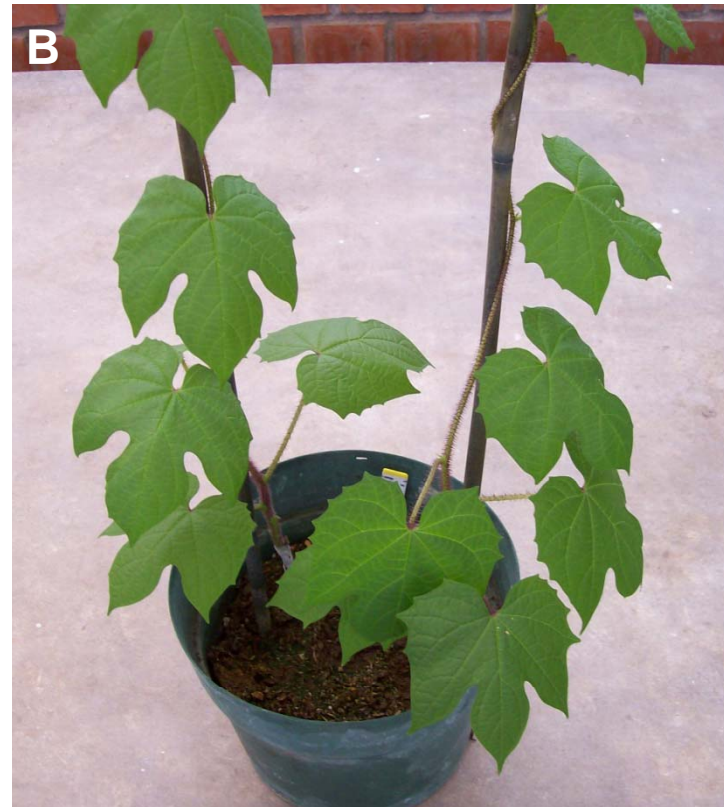




Somatic mutation



Clone CIP 401430 (Morada) showing somatic mutation (symptom of genetic distortion of leaves).



Indicator plant *I. seotsa* grafted with tissue scion from clone CIP 401430. No symptom is observed.

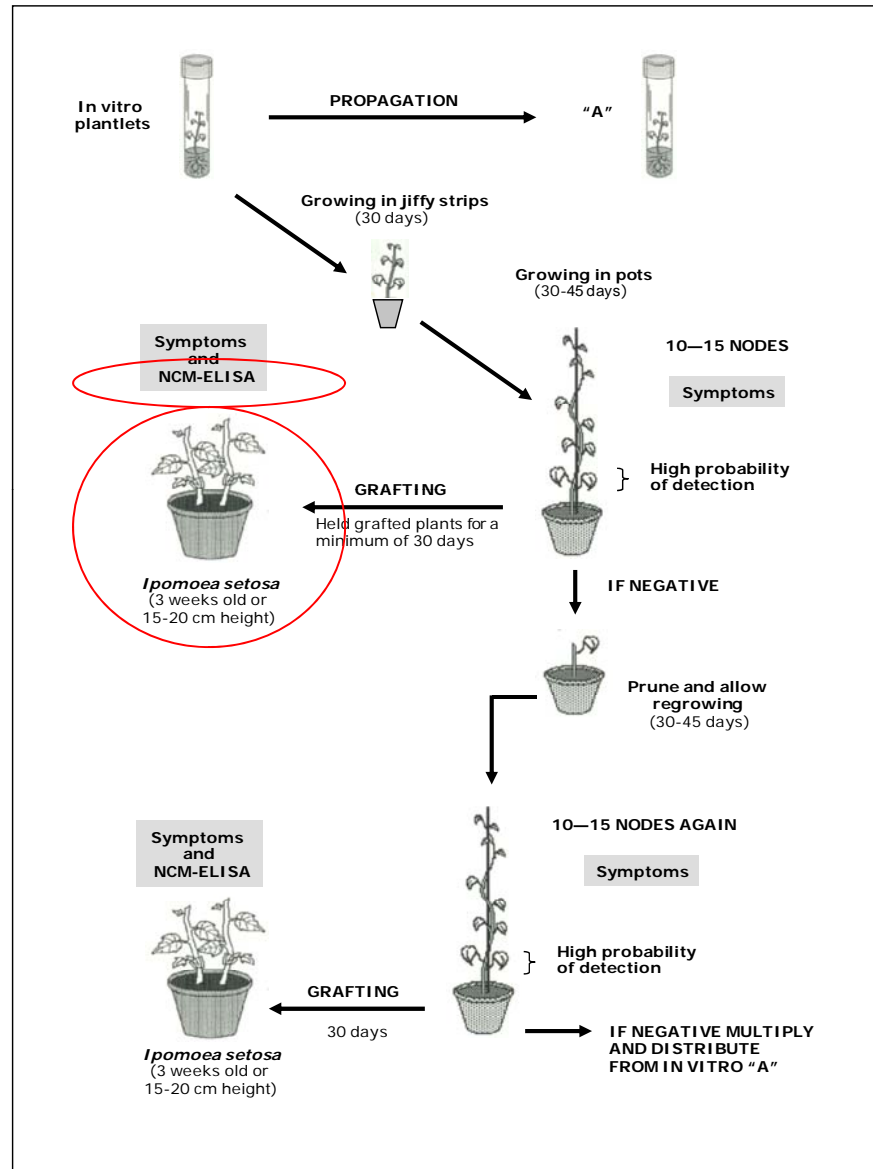


Final indexing report: Group 26 (G-26) 2010

| Record | Other code | Accession number | Labcode | Collecting number | Accession name | SP symptoms | <i>I. setosa</i> symptoms | PCR Serology | Results | Observations |
|--------|------------|------------------|---------|-------------------|------------------|-------------|---------------------------|---------------------|----------|------------------|
| 48 | 2 | 441443 | SG-4412 | IO1091 | Red Sweet Potato | | | | APPROVED | |
| 49 | 2 | 441129 | SG-4523 | IO1570 | So 10 | | C, RU | Begomo | REJECTED | |
| 50 | 2 | 420275 | SG-5417 | DLP 3311 | Zapallo | | Cp, Vc, Net, Vb | SPFMV, SPVG | REJECTED | |
| 51 | 3 | 102081.28 | SR-0190 | | | | C, RU | Begomo | REJECTED | |
| 52 | 3 | 189121.15 | SR-0192 | | | | | | APPROVED | SP (Variegation) |
| 53 | 1 | 189121.2 | SR-0193 | | | | | | APPROVED | |
| 54 | 5 | 189123.57 | SR-0199 | | | | | | APPROVED | |
| 55 | 5 | 189123.66 | SR-0200 | | | | | | APPROVED | |
| 56 | 5 | 189123.73 | SR-0202 | | | | | | APPROVED | |
| 57 | 1 | 189151.34 | SR-0214 | | | | | | APPROVED | |
| 58 | 5 | 194582.24 | SR-0253 | | | | | | APPROVED | |
| 59 | 4 | 194579.1 | SR-0255 | | | | | | APPROVED | |
| 60 | | 421081 | G-0001 | RCB IN- 1 | Huachano Chico | | | | APPROVED | |
| 61 | | 421089 | G-0021 | RCB IN- 21 | Quince Dias | | | | APPROVED | |
| 62 | | 420173 | G-0263 | DLP 318 | unknown | | | | APPROVED | |
| 63 | | 441741 | G-0343 | RCB IF- 48 | CAL-257 | | | | APPROVED | |
| 64 | | 422638 | G-0454 | UNPRG 25 | Morado De Huaura | | | | APPROVED | |
| 65 | | 400035 | G-0505 | RCB IF- 5 | IAC-85-17 | | | | APPROVED | |
| 66 | | 401493 | G-0604 | DLP 426 | | | C, Cp, RU, Vc | Begomo, SPFMV, SPVG | REJECTED | |
| 67 | | 401538 | G-0785 | DLP 1340 | Unknown | | | | APPROVED | |
| 68 | | 400460 | G-0874 | DLP 1792 | Blanca | | | | APPROVED | |
| 69 | | 400480 | G-0888 | DLP 1873 | unknown | | C, RU | Begomo | REJECTED | |
| 70 | | 400165 | G-1293 | DLP 1360 | unknown | | | | APPROVED | SP (Variegation) |
| 71 | | 400262 | G-1405 | DPW 2586 | Paulista | | | | APPROVED | SP (Variegation) |
| 72 | | 400414 | G-1768 | DLP 1734 | Leonarda | | | | APPROVED | |



Sweetpotato virus indexing procedure

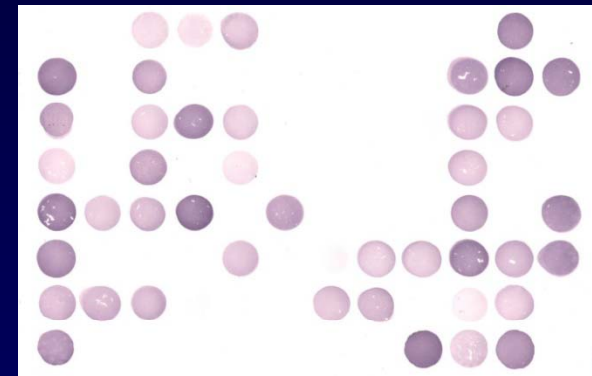


Indexing procedure for sweet potato viruses. NCM-ELISA is performed for 10 viruses (SPFMV, SPLV, SPVG, SPMSV, SPMMV, SPCSV, SPCFV, C-6, SPCV, and CMV). PCR is performed to detect begomoviruses.

Serological test



NCM-ELISA test



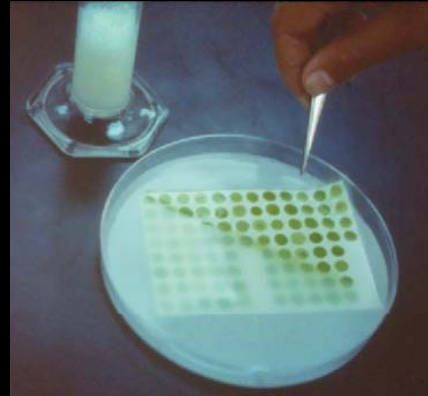
For 10 viruses

NCM-ELISA

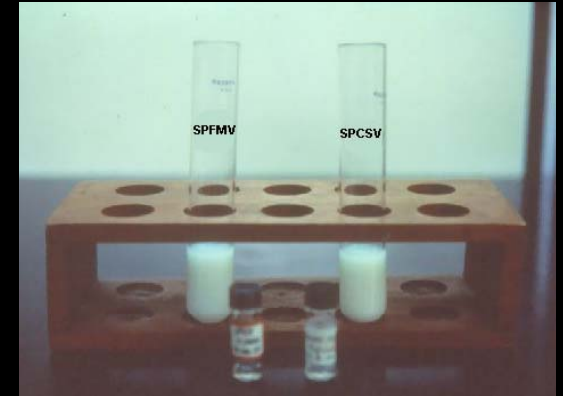
(A) Processing samples



(B) Blocking



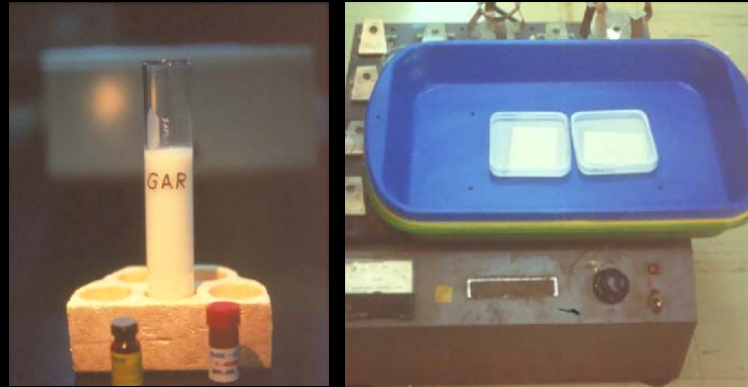
(C) 1st antibody



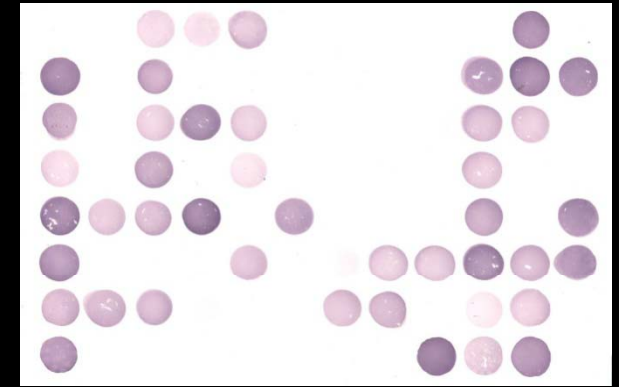
Washing



(D) 2nd antibodies (conjugate)

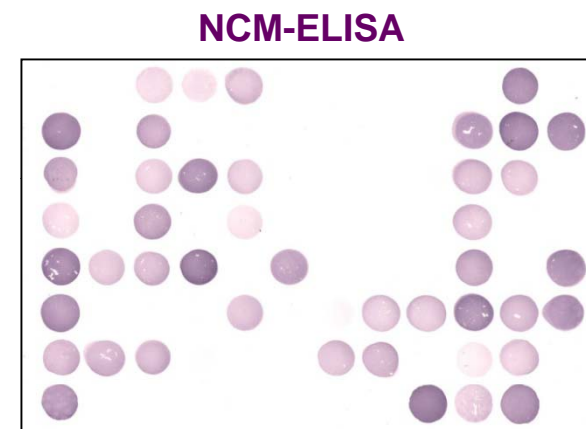


(E) Color development





Sweetpotato versus *I. setosa* versus serology



- Symptomless
- Low virus concentration

Indicator plant

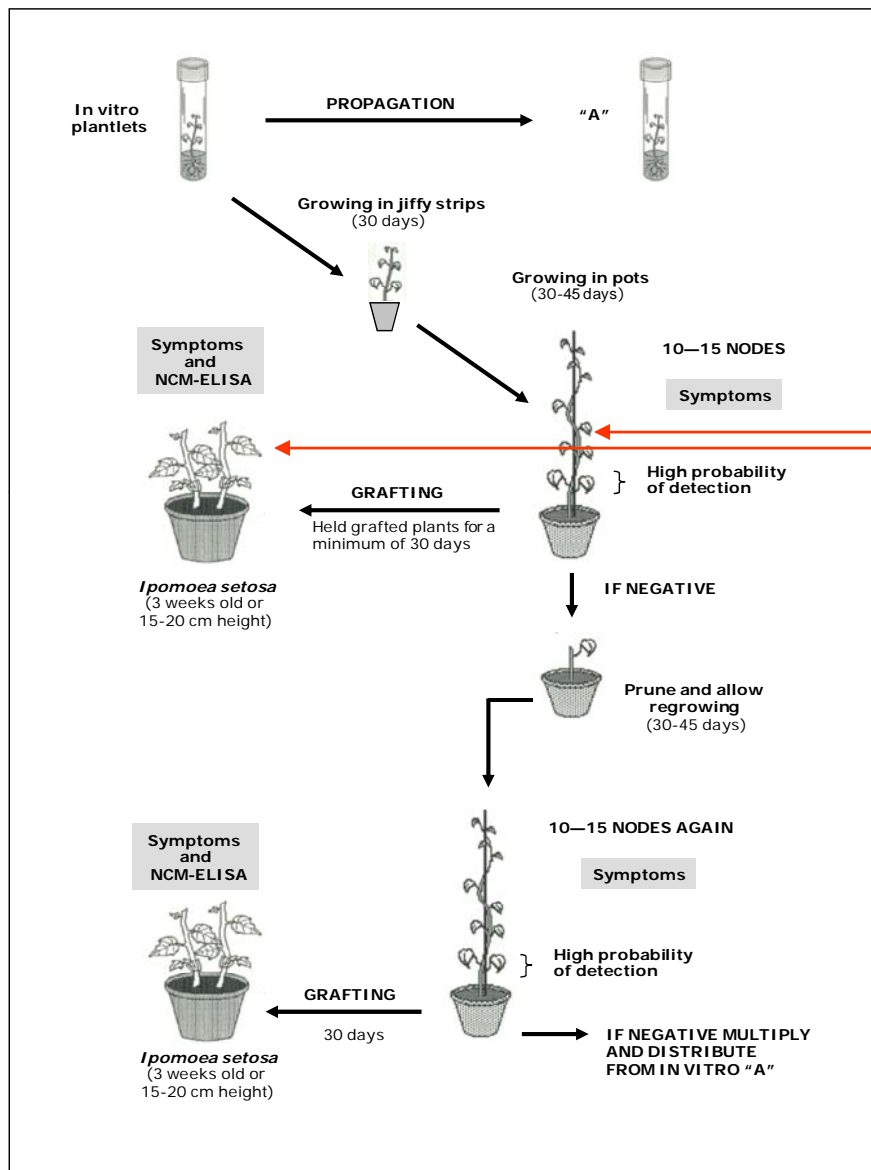
+

Serology (ELISA)

Reliable detection of viruses



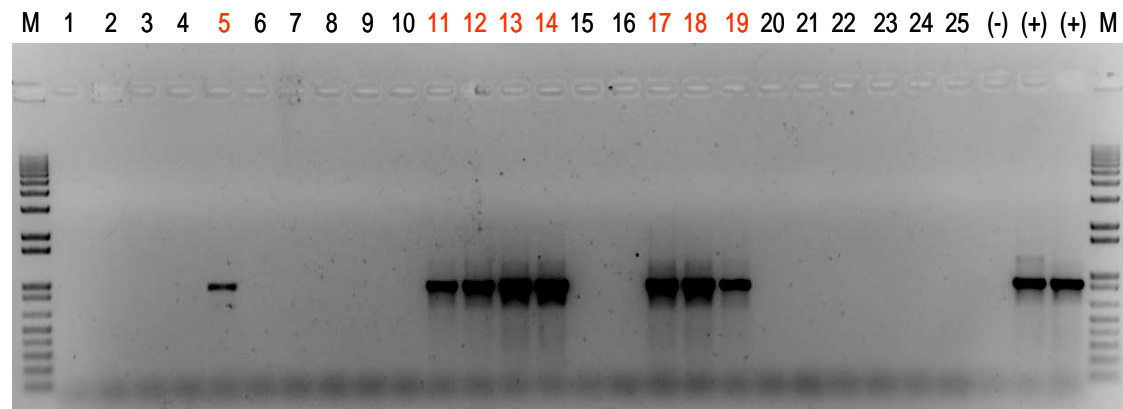
Sweetpotato virus indexing procedure



Indexing procedure for sweet potato viruses. NCM-ELISA is performed for 10 viruses (SPFMV, SPLV, SPVG, SPMSV, SPMMV, SPCSV, SPCFV, C-6, SPCV, and CMV). PCR is performed to detect begomoviruses.



Sweetpotato vs *Ipomoea setosa* vs PCR (begomovirus)



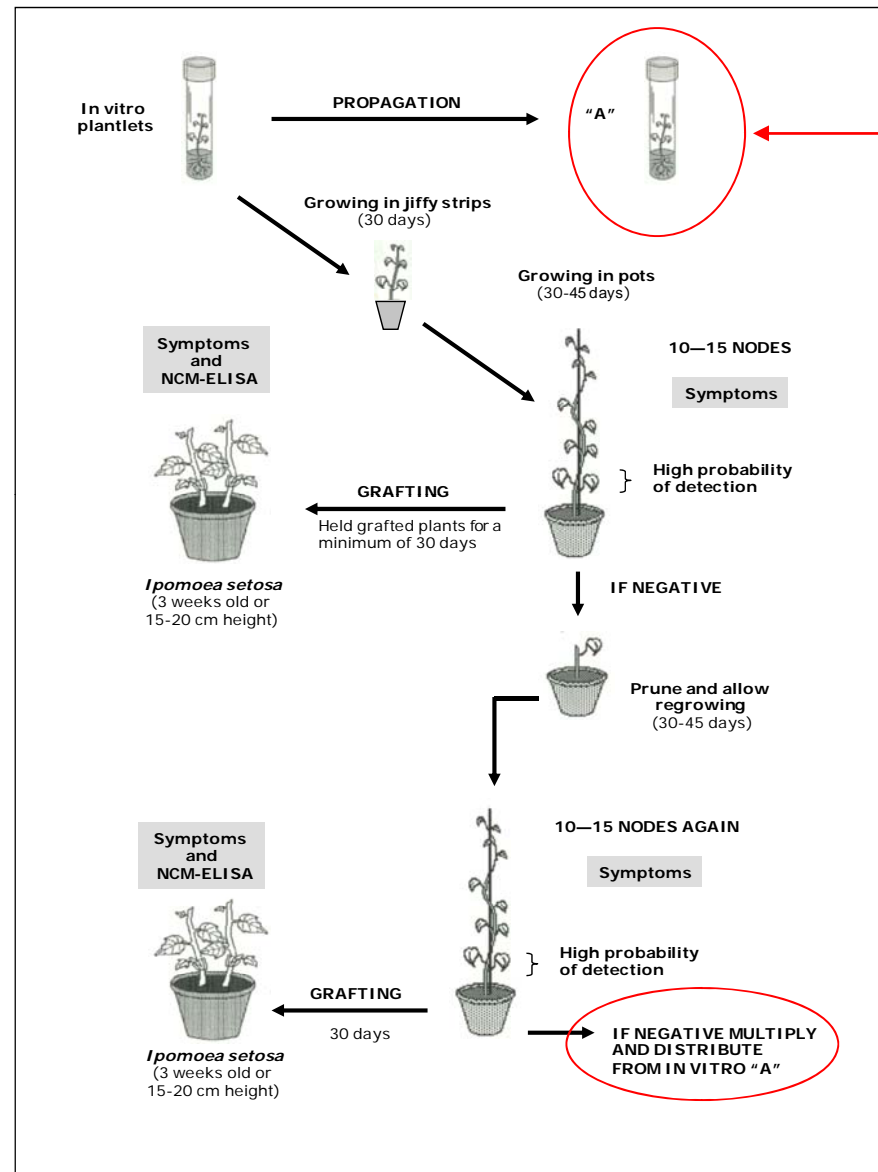
Li, R., Salih, S., and Hurtt, S. 2004. Detection of geminiviruses in sweetpotato by polymerase chain reaction. *Plant Dis.* 88:1347-1351.



45



Sweetpotato virus indexing procedure



Indexing procedure for sweet potato viruses. NCM-ELISA is performed for 10 viruses (SPFMV, SPLV, SPVG, SPMSV, SPMMV, SPCSV, SPCFV, C-6, SPCV, and CMV). PCR is performed to detect begomoviruses.



Final Results of Virus Indexing (Group 25, 2009)

| Record | Other code | Accession number | Labcode | Collecting number | Accession name | SP symptoms | <i>I. setosa</i> symptoms | Serology | PCR | Results | Observations |
|--------|------------|------------------|------------|-------------------|----------------|-----------------|---------------------------|------------|--------|----------|-----------------|
| 38 | 35 | 1 | CIP 401066 | SG-1477 | GTM 392 | Camote Morado | Ve,Vn | | | REJECTED | |
| 39 | 36 | 2 | CIP 401448 | SG-1524 | DLP 2888 | Morada | | | | APROVED | |
| 40 | 37 | 2 | CIP 403380 | SG-1558 | INIVIT 9 | Blanco Especial | | | | APROVED | |
| 41 | 38 | 2 | CIP 402666 | SG-1588 | INIVIT1049 | | RU,D,IVC | | Begomo | REJECTED | |
| 42 | 39 | 5 | CIP 402691 | SG-1592 | INIVIT1078 | | C,RU | | Begomo | REJECTED | |
| 43 | 40 | 5 | CIP 402721 | SG-1604 | INIVIT1216 | | RU | | Begomo | REJECTED | |
| 44 | 41 | 6 | CIP 401130 | SG-1691 | GTM 1100 | Camote Morado | IVC | | Begomo | REJECTED | |
| 45 | 42 | 7 | CIP 401375 | SG-1688 | SVG 27 | Six Weeks | D,IVC | | Begomo | REJECTED | |
| 46 | 43 | 4 | CIP 402619 | SG-1702 | INIVIT 217 | | RU | | Begomo | REJECTED | |
| 47 | 44 | 3 | CIP 402655 | SG-1717 | INIVIT1038 | | D,Ve,Vn,Ln | SPCaLV | | REJECTED | SPCaLV (PCR) |
| 48 | 45 | 5 | CIP 402696 | SG-1731 | INIVIT1084 | | C,IVC | | Begomo | REJECTED | |
| 49 | 46 | 1 | CIP 403435 | SG-1786 | CESDA861607 | 86-1607 | | | | APROVED | |
| 50 | 47 | 5 | CIP 400789 | SG-1790 | CENDA 20 | Jacobina 5 | RD,D,Ve,Vn,Ln | SPCaLV | Begomo | REJECTED | Virus C-9 (PCR) |
| 51 | 48 | 2 | CIP 401120 | SG-1816 | GTM 949 | Camote Blanco | | | | APROVED | |
| 52 | 49 | 5 | CIP 401277 | SG-1827 | DLP 3898 | Rojo-kinit | | | | APROVED | |
| 53 | 50 | 1 | CIP 420565 | SG-1852 | DLP 66 | Espelma | D,Ve | SPFMV,SPVG | | REJECTED | |
| 54 | 51 | 6 | CIP 420163 | SG-1859 | DLP 123 | Morado Morado | | | | APROVED | |
| 55 | 52 | 2 | CIP 420181 | SG-1907 | DLP 887 | unknown | Cp | SPFMV,SPVG | | REJECTED | |
| 56 | 53 | 6 | CIP 420715 | SG-1935 | DLP 1312 | Zapallona | RD,Ve | SPFMV,SPVG | Begomo | REJECTED | |
| 57 | 54 | 6 | CIP 420291 | SG-2089 | DLP 3549 | unknown | | | | APROVED | |
| 58 | 55 | 5 | CIP 420113 | SG-2154 | ARB 323 | Camote Papa | Cp | SPFMV,SPVG | | REJECTED | |
| 59 | 56 | 1 | CIP 420114 | SG-2156 | ARB 347 | Camote | | | | APROVED | |
| 60 | 57 | 6 | CIP 420488 | SG-2172 | ARB 425 | Espelma | | | | APROVED | |
| 61 | 58 | 3 | CIP 442115 | SG-2283 | IITA-CL 57 | | C,RU,D | | Begomo | REJECTED | |
| 62 | 59 | 2 | CIP 443083 | SG-2384 | IITA-CL 249 | TIS 2180 | C,B | | Begomo | REJECTED | |
| 63 | 60 | 3 | CIP 442858 | SG-2452 | IITA-CL 377 | TIS 8030 | | | | APROVED | |



Virology Lab Info Management System (CIPVIR)

Maintenance ▶ Register a Diagnostic ▶ Distribution KITS ▶ Reports ▶ Tools ▶ Help ▶



Welcome!

The 'Virology Lab Info Management System' provides services that the 'Virology Section' of the 'Crop Protection' provides to the CIP staff and external clients.

The facilities are: Register the request of a diagnostic (Range), register the results of diagnostics, maintain on pathogens, etc, print reports by different criteria.

We welcome any comments and/or suggestions.
RIU@cgiar.org

Virology Lab Info Management System version 1.0 (2003)+++Last software review: April, 2003+++Contact: CIP-RIU@cgiar.org

G-32B

<http://sol/appdb/research/Div4ICM/CIPVIR/diagnosticIndicatorPlant.aspx?strYear=2011&strNumber=5>

<http://intranet.cip.cgiar.org/appdb/research/Div4ICM/CIPVIR/diagnosticIndicatorPlant.aspx?strYear=2011&strNumber=5>

<http://sol/appdb/research/Div4ICM/CIPVIR/diagnosticNCMElisa.aspx?strYear=2011&strNumber=8>

<http://intranet.cip.cgiar.org/appdb/research/Div4ICM/CIPVIR/diagnosticNCMElisa.aspx?strYear=2011&strNumber=8>



Outline

- Introduction
- ISO/IEC 17025
- Virus indexing procedure
- **Analysis of indexing results**

Number of sweetpotato accessions indexed for viruses between years 2006 and 2011

| Year | Group | Analyzed accessions | Positive accessions | By grafting (<i>I. setosa</i>) | By serology (NCM- ELISA) | By PCR (Begomovirus) |
|--------------|-------|---------------------|------------------------|-------------------------------------|--|--------------------------|
| 2011 | 34 | 232 | 85 | 81 | 43 | 49 |
| | 33 | 11 | 9 | 9 | 5 | 7 |
| | 32 | 163 | 82 | 72 | 45 | 49 |
| | 31 | 244 | 92 | 89 | 47 | 58 |
| 2010 | 30 | 131 | 57 | 55 | 38 | 24 |
| | 29 | 220 | 86 | 86 | 44 | 26 |
| | 28 | 66 | 2 | 1 | 1 | 0 |
| | 27 | 243 | 123 | 122 | 103 | 62 |
| 2009 | 26 | 329 | 141 | 141 | 46 | 103 |
| | 25 | 231 | 103 | 103 | 91 | 59 |
| 2008 | 24 | 201 | 77 | 74 | 31 | |
| | 23 | 72 | 15 | 10 | 15 | |
| | 22 | 272 | 111 | 111 | 43 | |
| 2007 | 21 | 197 | 100 | 100 | 9 | |
| | 20 | 10 | 0 | 0 | 0 | |
| | 19 | 151 | 44 | 44 | 9 | 2 |
| 2006 | 18 | 193 | 60 | 60 | 4 | |
| | 17 | 65 | 40 | 40 | 32 | |
| | 16 | 318 | 114 | 112 | 21 | |
| TOTAL | | 3,349 | 1,341 (40%) | 1,310 | 627 | Begomovirus = 439 |
| | | | | | SPFMV= 361 SPVG= 270 SPCV= 72 SPCSV= 52 SPCFV= 35 SPLV= 9 SPMSV= 4 C-6= 1 CMV= 1 | |

Country of origin of accessions which tested positively to viruses during indexing

| Country | NCM-ELISA | | | | | | | PCR | | <i>I. setosa</i> (symptoms) | | | |
|------------------|-----------|------|-------|------------|-----|------------|-------------|-------------|--------------|-----------------------------|------------------------------|-----------|---------|
| | Potyvirus | | | Carlavirus | | Crinivirus | Cavemovirus | Begomovirus | Solendovirus | Begomovirus | Cavemovirus/ Solendovirus | Potyvirus | Other ? |
| | SPFMV | SPVG | SPMSV | SPCFV | C-6 | SPCSV | SPCV | | SPVCV | | | | |
| Argentina | | | | | | | | 4 | | 9 | | | 1 |
| Bolivia | | | | | | | | | | 5 | | 1 | |
| Brazil | | | | | | | | 2 | | 5 | | 1 | 2 |
| China | | | | | | | | | | 2 | | | |
| Colombia | | | 1 | | | | | 3 | | 7 | | | 1 |
| Costa Rica | | | | | | | | 2 | | | | | 1 |
| Cuba | 1 | | | | | | 3* | 15 | | 34 | 7 | | 2 |
| Dominican Repub. | | | | | | | 1 | 8 | 1 | 64 | 10 | 1 | |
| Ecuador | | | | | | | | 3 | | 5 | 1 | 2 | |
| Guatemala | | | | | | | 6 | 5 | 2 | 9 | 10 | | |
| Honduras | | | | | | | | | | 1 | | | |
| India | 10 | 6 | | | | | 3 | 14 | | | 2 | | |
| Jamaica | | | | | | | | 1 | | | 8 | 1 | |
| Japan | 1 | 1 | | | | | | 2 | | 2 | | 1 | 2 |
| Sri Lanka | | | | | | | | | | 1 | | | |
| Mexico | | | | 3 | | | 1+1 | 2 | | 5 | | | |
| Malaysia | | | | | | | | | | | | | 1 |
| Nicaragua | | | | | | | | | | | | | 1 |
| New caledonia | | | | | | | | 1 | | 1 | | | |
| Nigeria | 1 | 1 | | | | | | 4 | | 48 | | | |
| Panama | | | | | | | 1* | 5 | | 2 | | | |
| Peru | 135 | 99 | 2 | 12 | | 5 | | 12 | | 50 | 3 | 4 | 4 |
| Philippines | | | | | | | | | | 1 | | | |
| Papua New Guinea | 44 | 25 | | 1 | | | 7 | 8 | | 58 | 3 | 1 | 6 |
| Puerto Rico | 1 | 1 | | | | | | 2 | | 1 | 1 | 1 | 1 |
| Paraguay | 2 | 2 | | | | | | | | 3 | | | |
| Solomon Islands | 2 | 1 | | 1 | | | 1 | 4 | | 6 | | | 1 |
| Thailand | | | | | | 1 | | | | 6 | | | 1 |
| Tonga | | | | | | | | | | 2 | | | |
| Taiwan | 6 | 3 | | | | 1 | 2 | 6 | | 3 | | | 2 |
| Uganda | | | | | | | | 1 | | | | | |
| USA | 1 | 1 | | | 1 | | 3* | 4 | | 7 | 1 | | |
| Saint Vincent | | | | | | | | 1 | | 5 | | | |
| Venezuela | | | | | | | | 1 | | 4 | 4 | 2 | 1 |



Outline

- Introduction
- ISO/IEC 17025
- Virus indexing procedure
- Analysis of indexing results
- **Research derived from indexing**



Isolate C-9 (CIP 400851)

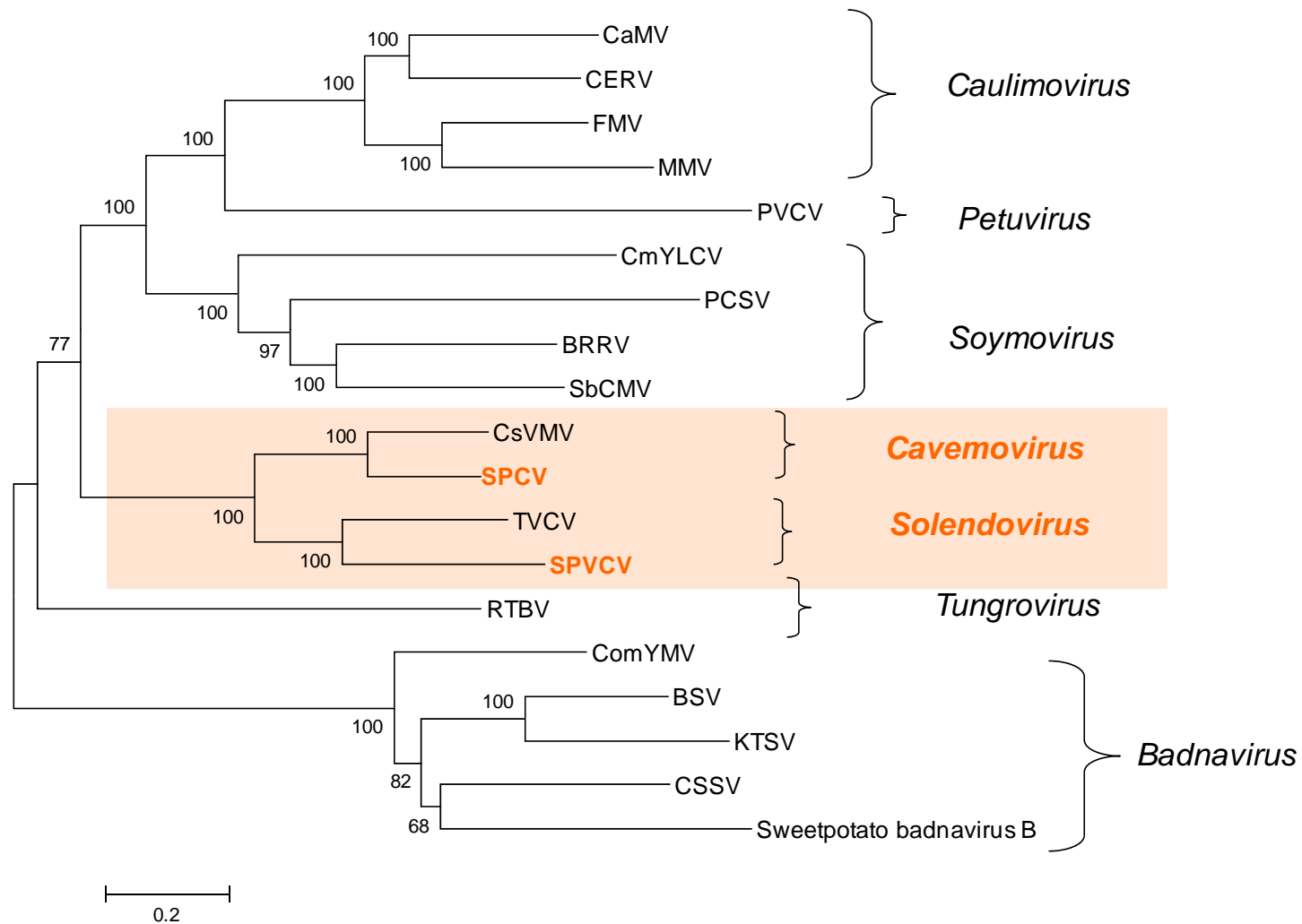
Dominican Republic, Sosa 30, “Chambrita” cultivar



2002



Phylogenetic analysis (complete genome)



J. De Souza's Thesis

De Souza y Cuellar. 2011. Archives of Virology 156: 535-537.
Cuellar et al 2011. Journal of General Virology 92: 1233-1243.
Geering et al 2009. Archives of Virology 155: 123-131.



Geographic distribution

Based on serological and PCR test, respectively



J. De Souza's Thesis



Identification of begomovirus isolates

M. Galvez's Thesis



**San Vicente
(400025)**



**Perú-6
(421076)**



**Perú-10
(420993)**



**Jamaica-12
(401201)**



**Cuba-5
(402645)**

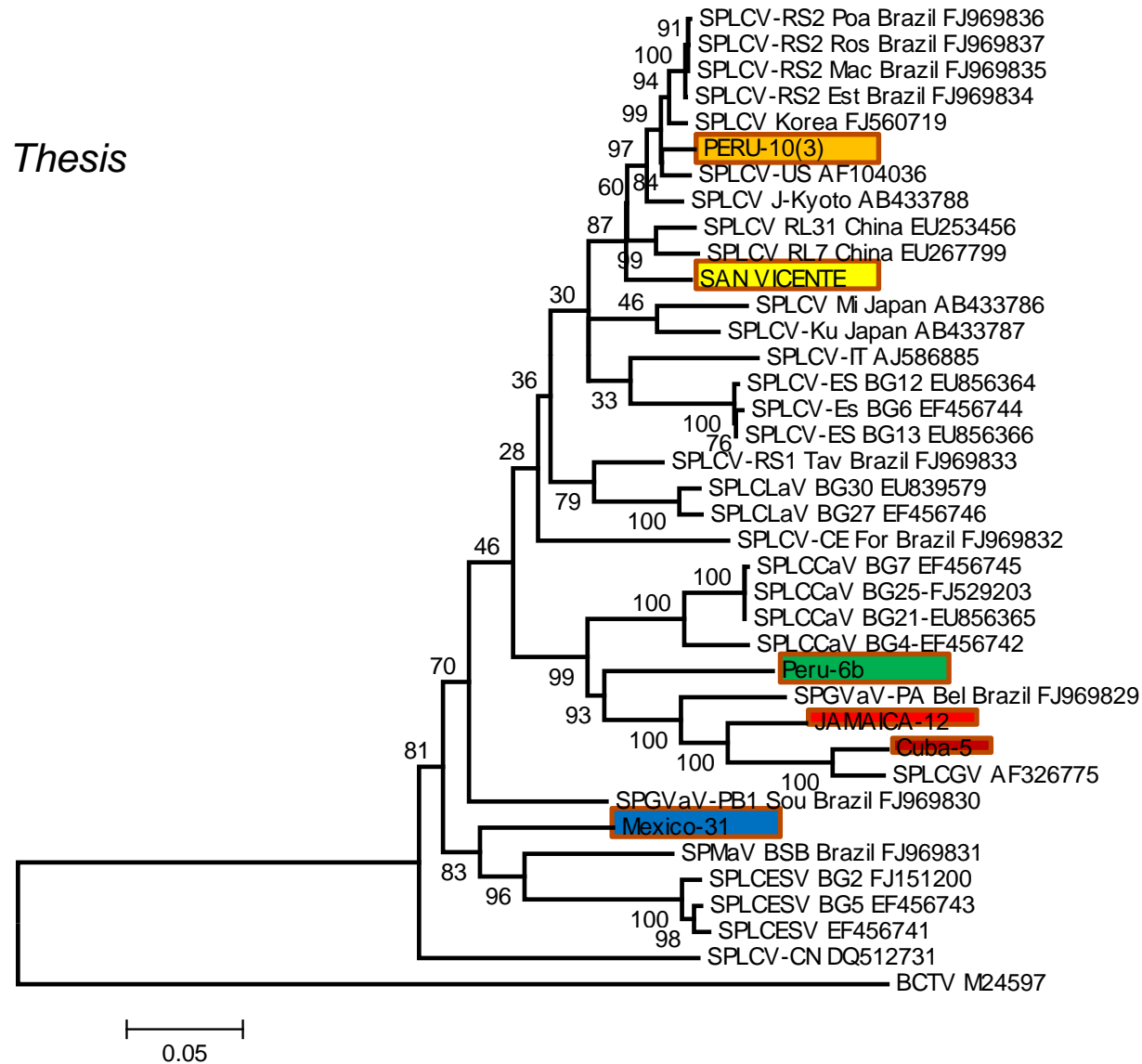


**México-31
(401468)**



Swepovirus phylogenetic analysis (complete genomes)

M. Galvez's Thesis





New virus?



CIP 440962 (Papua New Guinea)



CIP 420883 (Peru)



New virus?



CIP 401580 (Guatemala)



New virus?



CIP 400789 (Dominican Rep.)



Outline

- Introduction
- ISO/IEC 17025
- Virus indexing (SP in vitro germplasm)
- Analysis of indexing results
- Research derived from indexing
- **Maintaining ISO accreditation**



External Audit conducted by UKAS (C. Jeffries), Octobre 2009



ISO 17025



Testing laboratory
No. 4229



Equipments



Digital balance



Rotatory shaker



pHmeter



Magnetic stirrer



Freezer



Equipment calibration



Digital balance



Refrigerator



Monitoring temperature



Manual temperature recording

Data logger with thermocouple

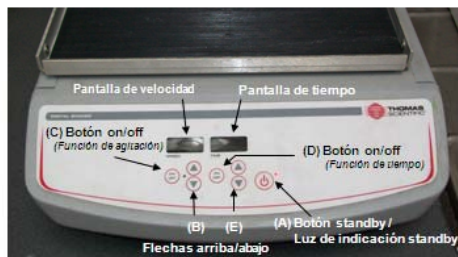


Equipment operational instruction

IS - HQU - 0304010167

INSTRUCTIVO DE USO DE AGITADOR ORBITAL DIGITAL

MARCA: THOMAS SCIENTIFIC
MODELO: 980224
CODIGO CIP: 0304010167



Poniendo operativo el equipo

1. Presionar el botón standby (A) para mover la unidad del modo standby.

Seleccionando la velocidad de agitación

2. Presionar la flecha arriba/abajo (B) hasta alcanzar la velocidad deseada.
3. Presionar el botón on/off (C) para comenzar la función de agitación.
4. Ajustar la velocidad con las flechas arriba/abajo (B) sin interrumpir la agitación.
5. Parar la función de agitación presionando el botón on/off (C).

Seleccionando el tiempo a cero (0:00) y en modo continuo (tiempo acumulado)

6. Presionar y mantener presionado el botón on/off (D).
7. Presionar simultáneamente ambas flechas arriba y abajo (E) para colocar el tiempo a cero.
8. Presionar el botón on/off (D). La pantalla de tiempo indicará el tiempo acumulado.
9. Para parar el tiempo, presionar nuevamente el botón on/off (D).

Programando el tiempo

10. Presionar las flechas arriba/abajo (E) hasta alcanzar el tiempo deseado.
11. Comenzar esta función presionando el botón on/off (D).
12. Para interrumpir el ciclo de tiempo automático antes que se complete, presionar el botón on/off (D).

Apagando la unidad

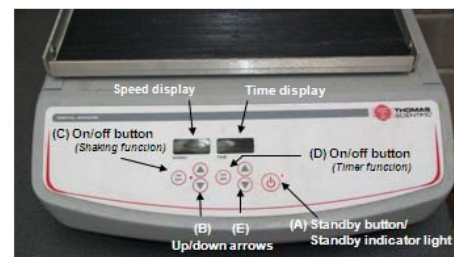
13. Presionar el botón standby (A) para mover la unidad al modo standby.

1 of 2

IS - HQU - 0304010167

INSTRUCTION SHEET TO USE DIGITAL ORBITAL SHAKER

SUPPLIER: THOMAS SCIENTIFIC
MODEL: 980224
CIP CODE: 0304010167



Getting ready

1. Press the standby button (A) to move the unit from standby mode.

Setting speed

2. Press the up/down arrow (B) until you reach the desired speed.
3. Press the on/off button (C) to start the shaking function.
4. Adjust speed without interrupting shaking by using the up/down arrows (B).
5. Stop the shaking function by pressing the on/off button (C).

Setting time to zero (0:00) and continuous mode: Accumulated time

6. Press and hold the on/off button (D).
7. Set time to zero by simultaneously pressing both the up and the down arrows (E).
8. Press the on/off button (D). The display will indicate accumulated time.
9. Stop timer by pressing the on/off button (D) again.

Setting timed mode: Programmed time

10. Press the up/down arrows (E) until you reach the desired time.
11. Start this function by pressing the on/off button (D).
12. To interrupt an automatic timing cycle before it is completed, press the on/off button (D).

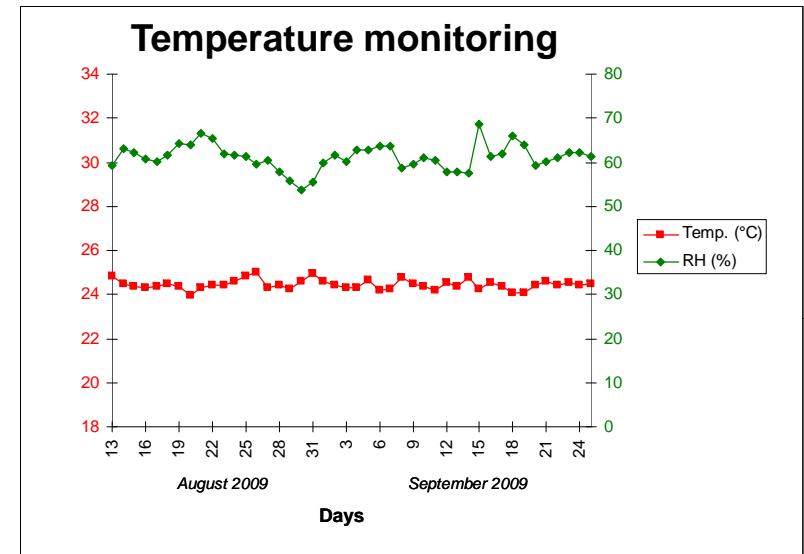
Turning unit off

13. Press the standby button (A) to move the unit to standby mode.

2 of 2



Control of all factors affecting reliability on virus detection



- Virus variability
- Low virus concentration on the SP plants
- Viruses no uniformly distributed on the SP plants
- Symptom expression on grafted *I. setosa* plants

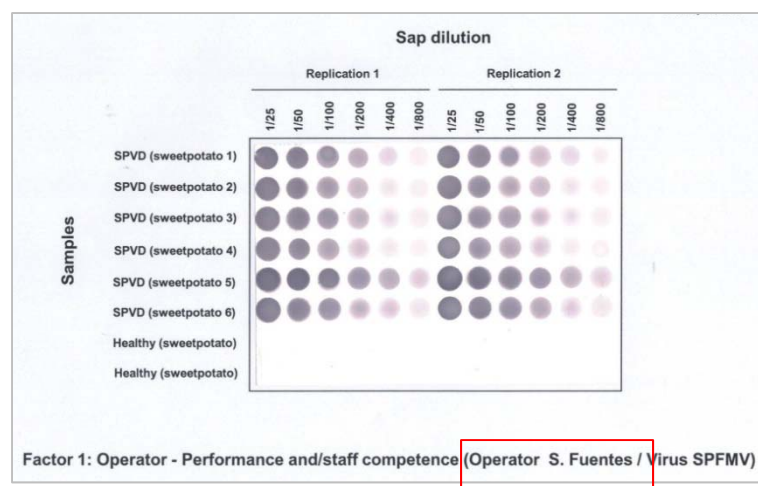
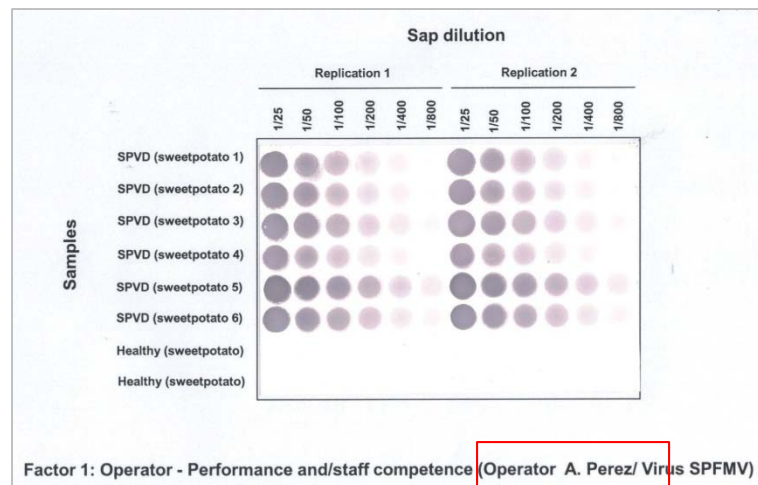


Personnel: competent / trained





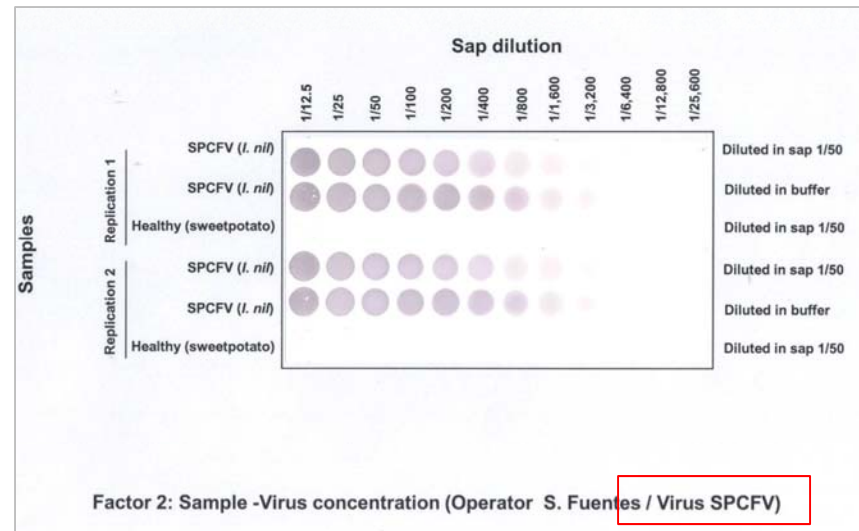
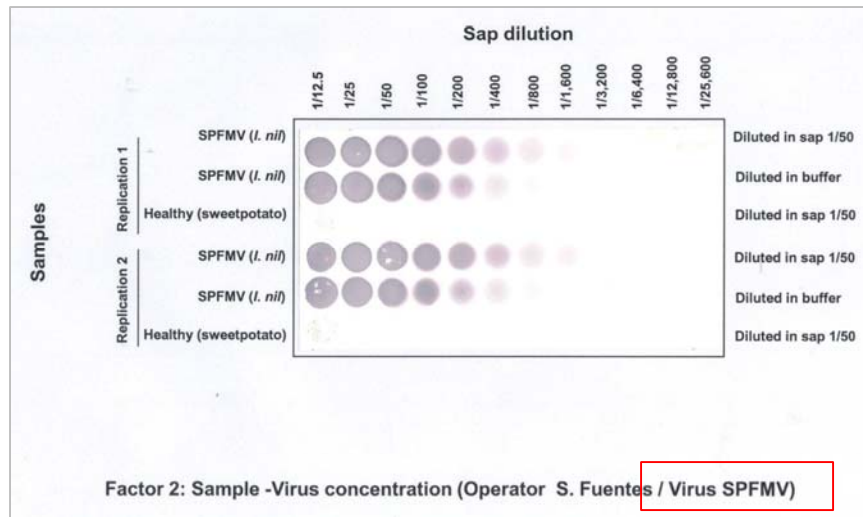
February 28, 2012





IQC / Uncertainty of measurement or NCM-ELISA

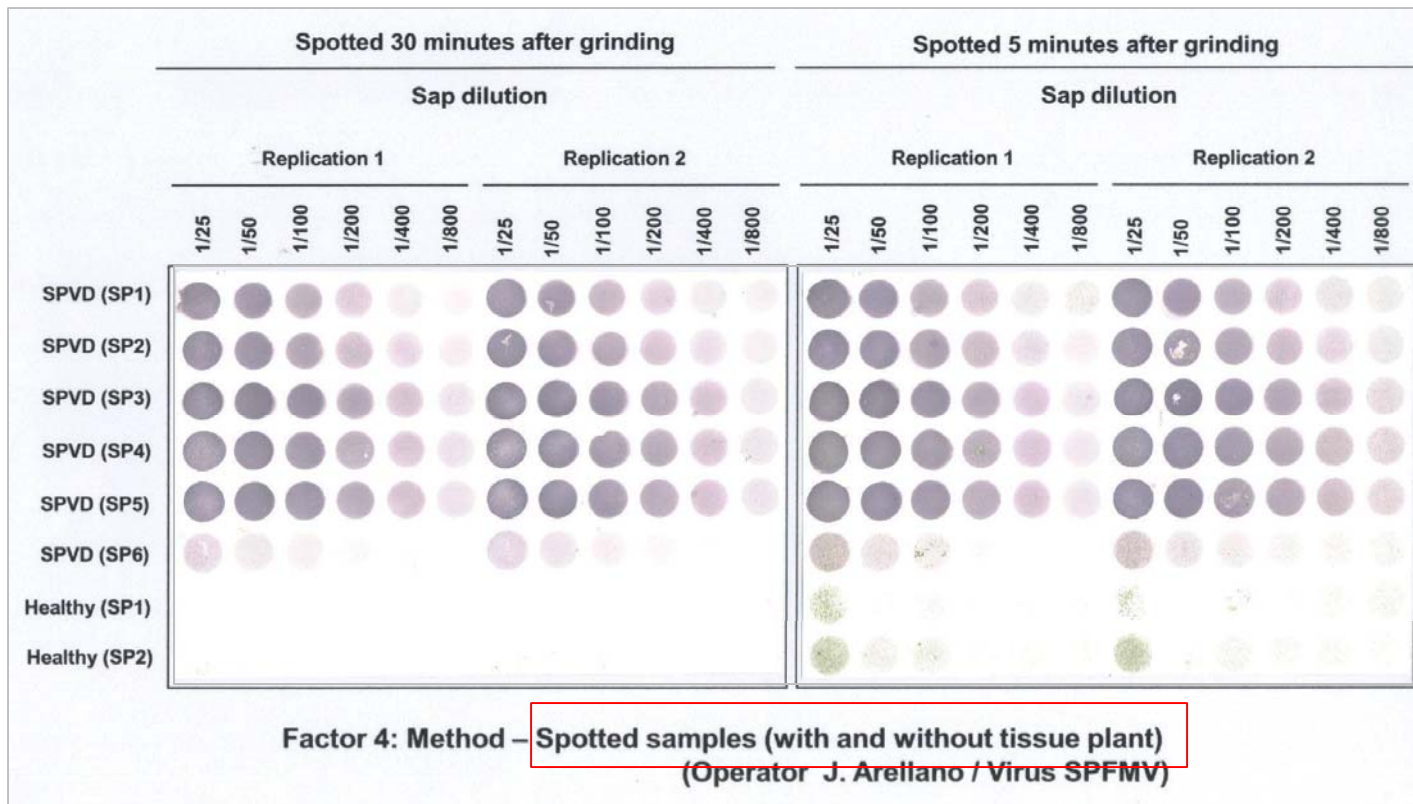
February 28, 2012





IQC / Uncertainty of measurement or NCM-ELISA

March 05, 2012





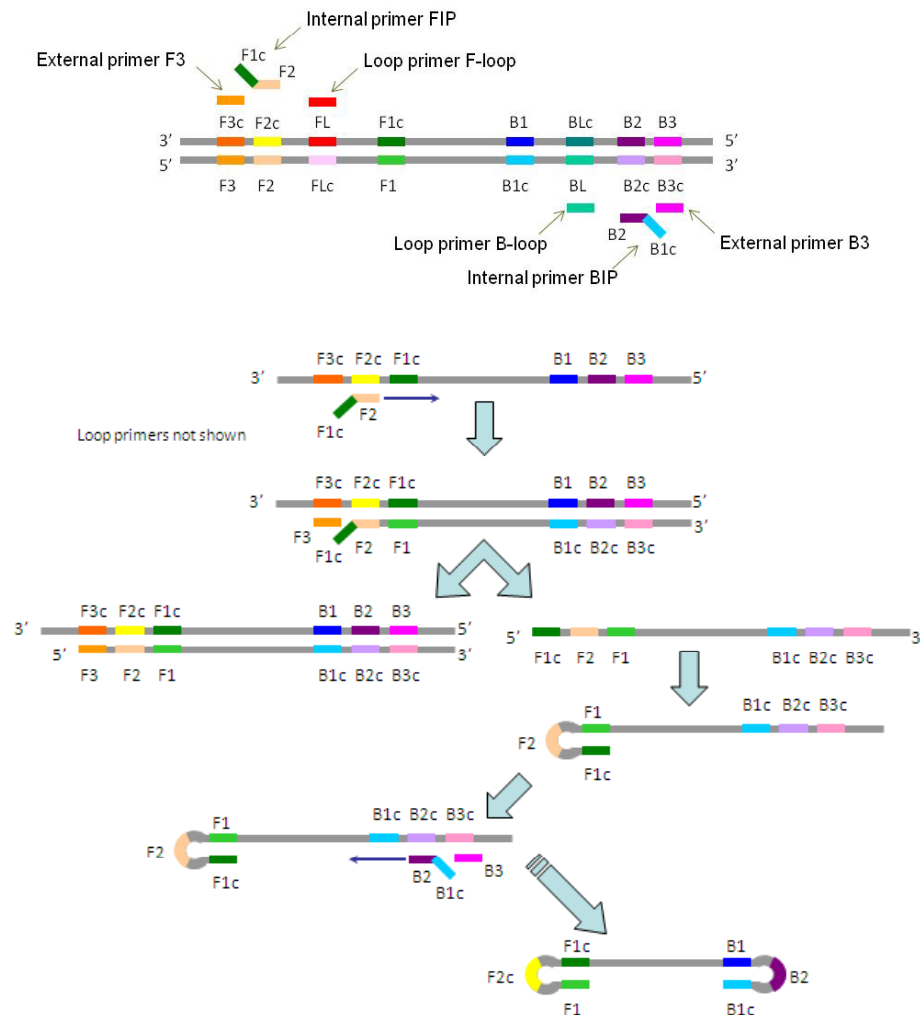
Outline

- Introduction
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- Analysis of indexing results
- Research derived from indexing
- Maintaining ISO accreditation
- **New molecular tests (validation)**



New molecular test (LAMP)

LAMP: Loop-mediated isothermal AMPlification

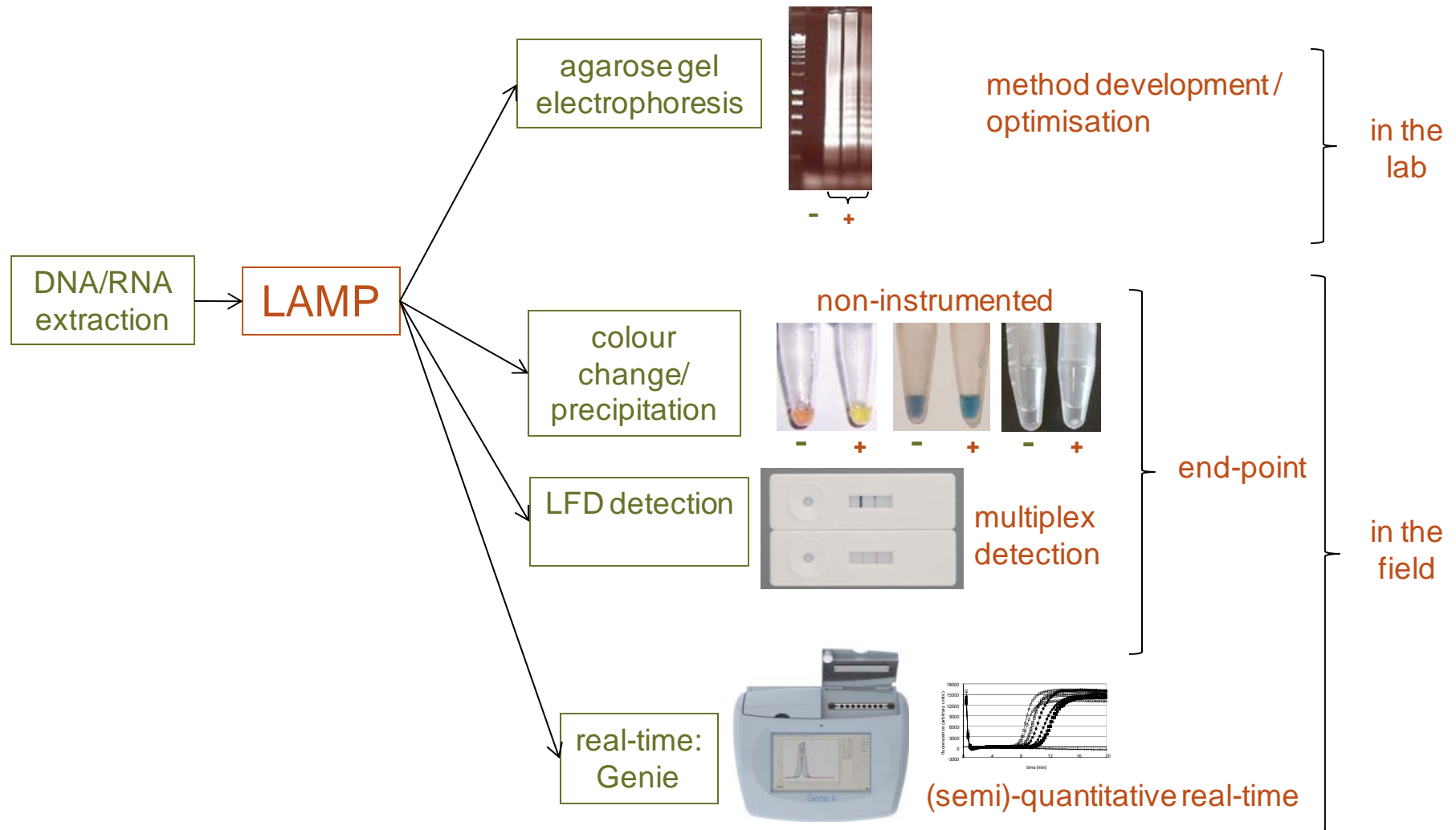


- Non-denatured target
- Isothermal reaction at 65°C
- 20-60 minute reaction time

Overcoming many of the limitations of PCR-based methods (cost and complexity of thermal cycling equipment)



Detection of LAMP products



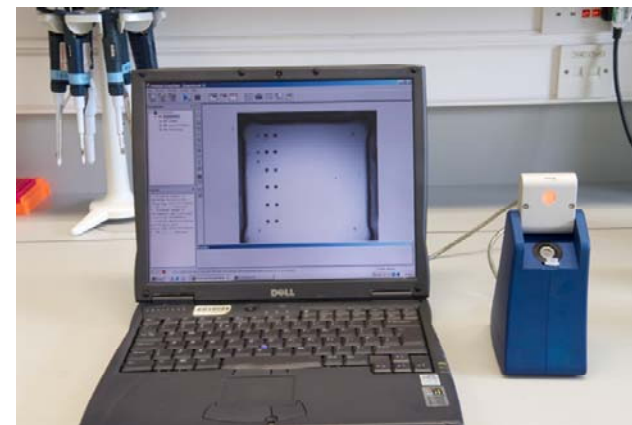
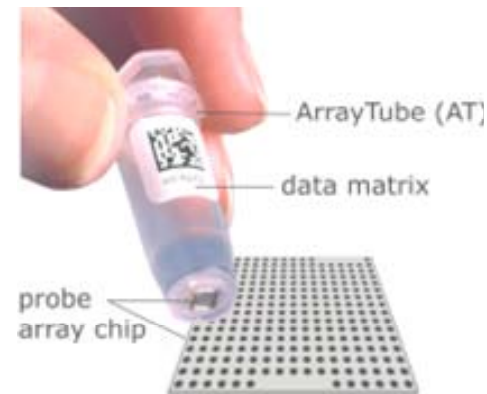


New technology (ClonDiag array)

Array Tube

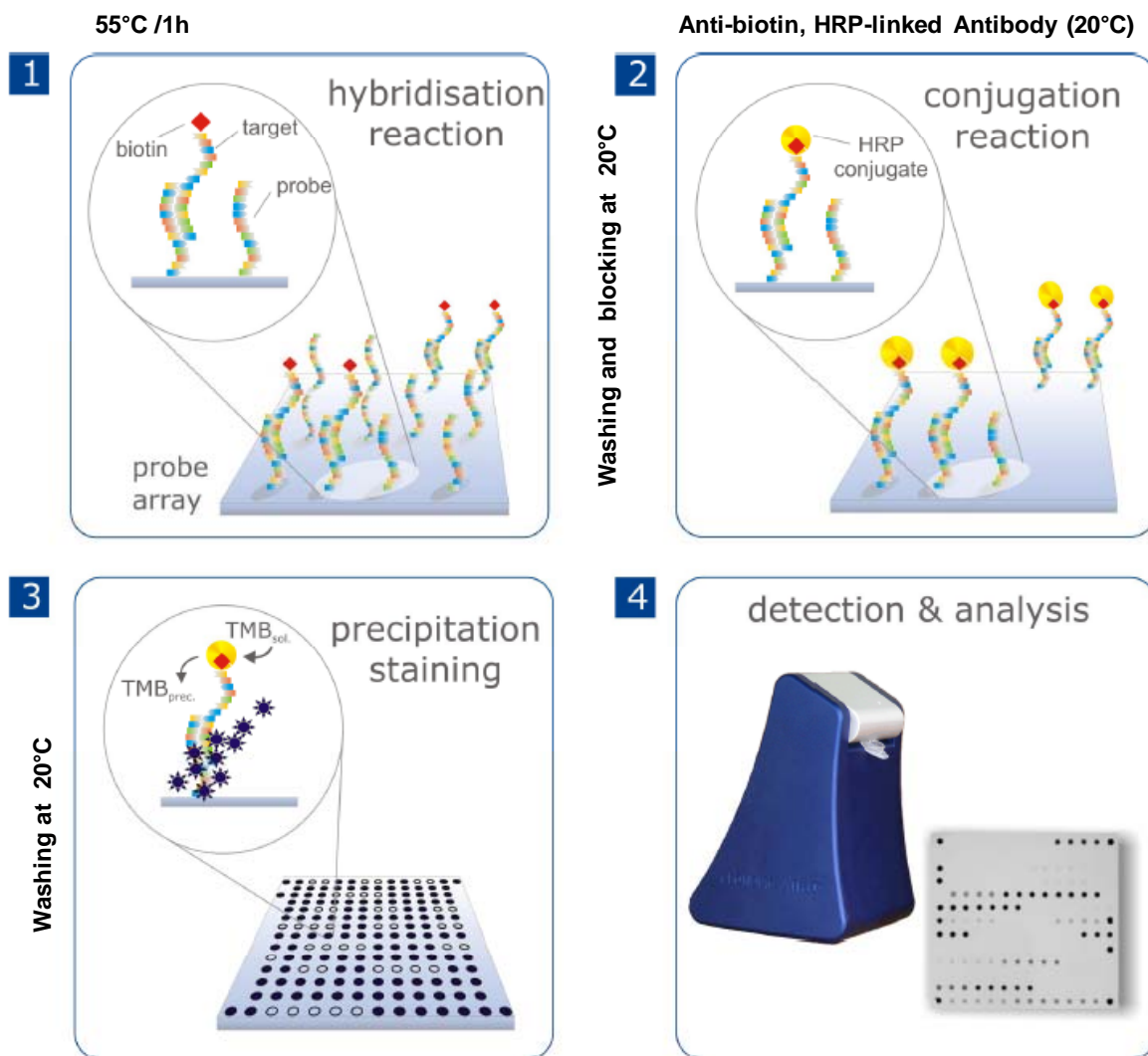
- Mini microarray embedded in tube
- Up to 80 features
- One step labelling
(biotin amp)
- Cheap scanner
- Manipulations in tube

CLONDIAG





Clondia Array Work Flow



Tetramethyl benzidine (TMB) or o-Dianisidine

When using array tube

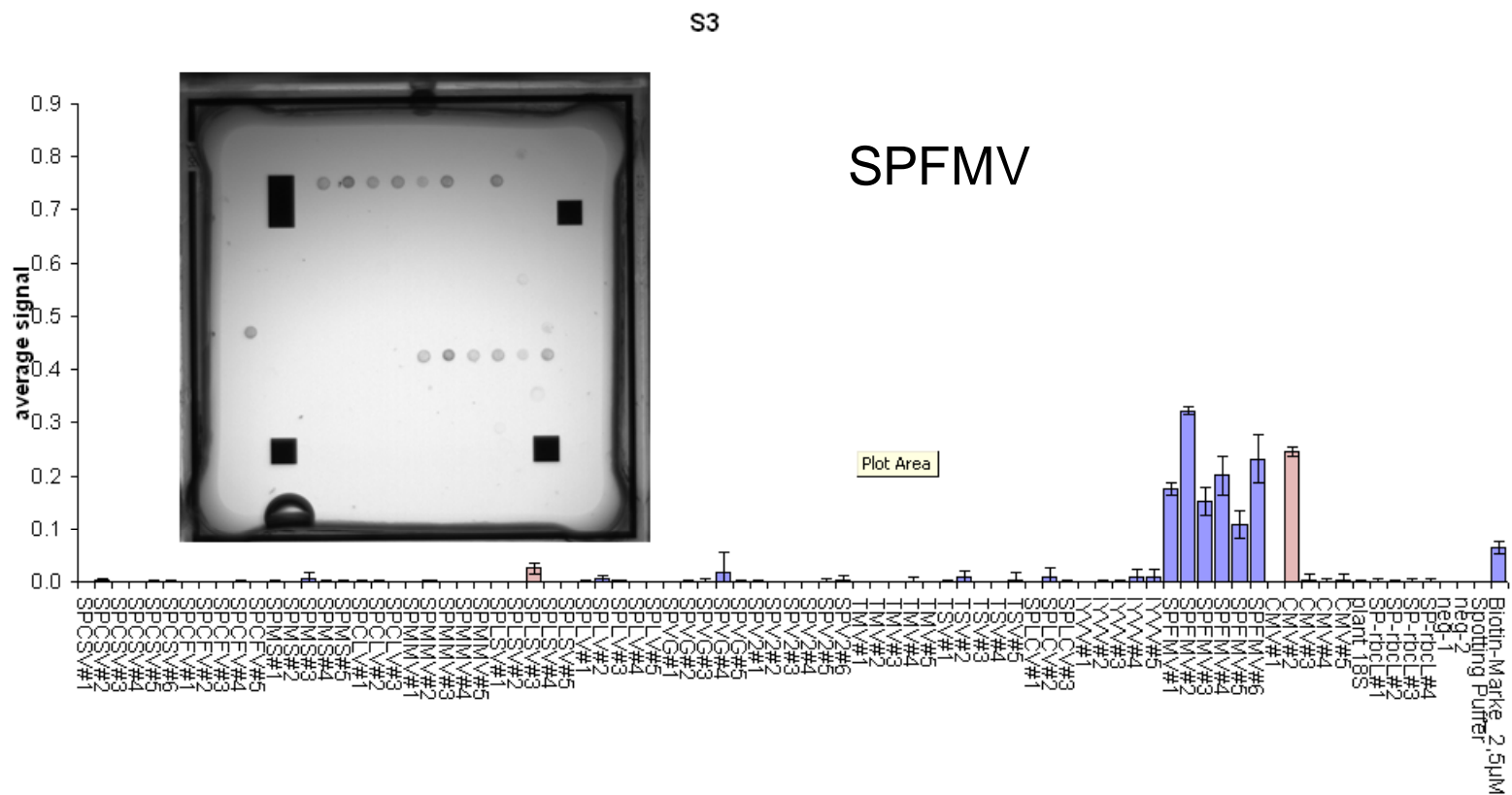
- Do not allow dry out
- Never spin



www.clondia.com/



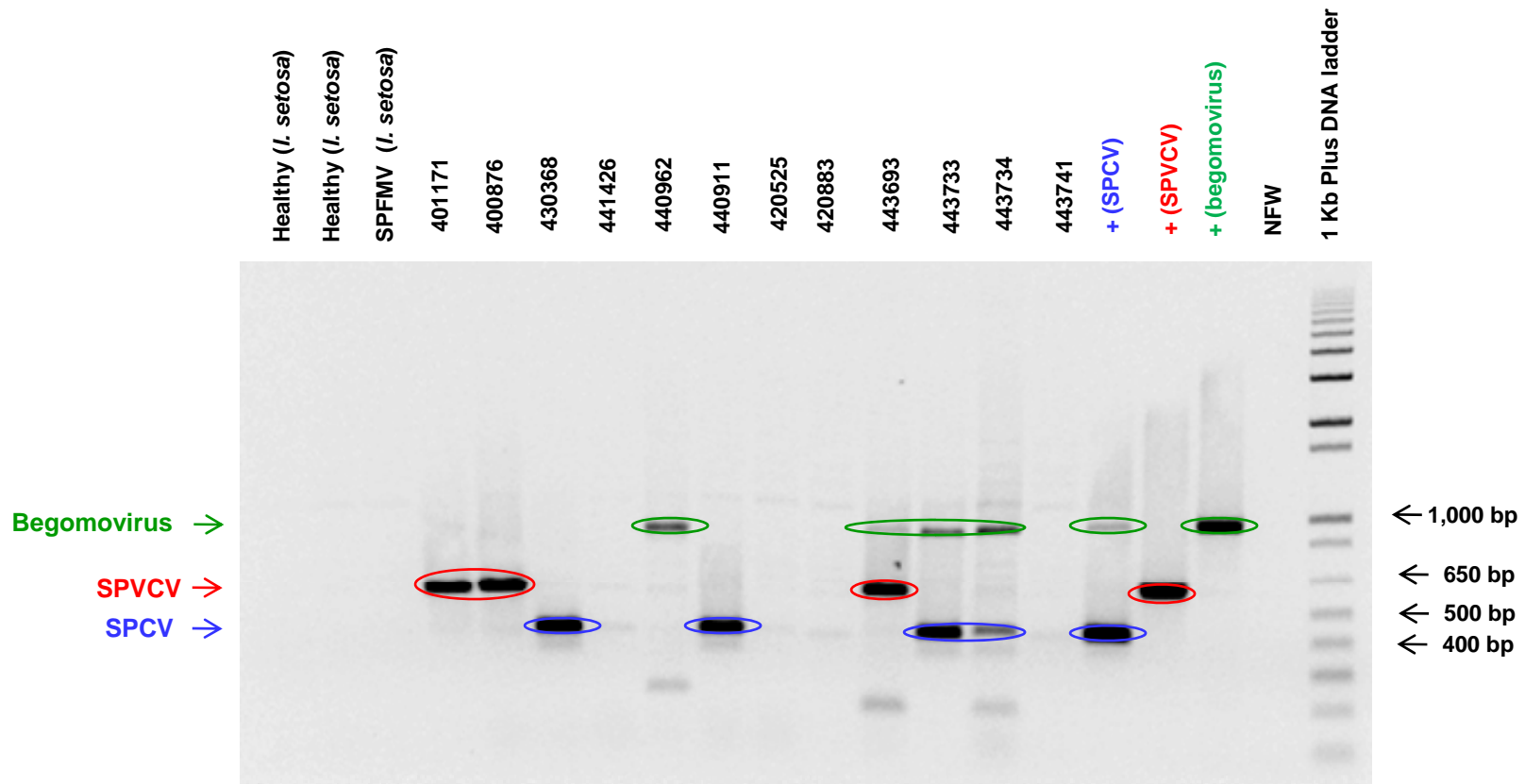
Sweetpotato Virus Array





Multiplex PCR (under validation)

For detecting DNA viruses: SPCV, SPVCV, begomovirus



A. Berrocal's Thesis



Staff involved in SP virus indexing

- Segundo Fuentes
 - Jaime Arellano
 - Marco Meza
 - Ana Pérez
 - Raúl Lara
 - Brenda Zea
 - Juan Cruzado
 - Miguel Velarde
 - Edwin Rojas
 - D. Hiraoka
- HQU
- IVU
- RIU



CIP's ISO Team 2011

