



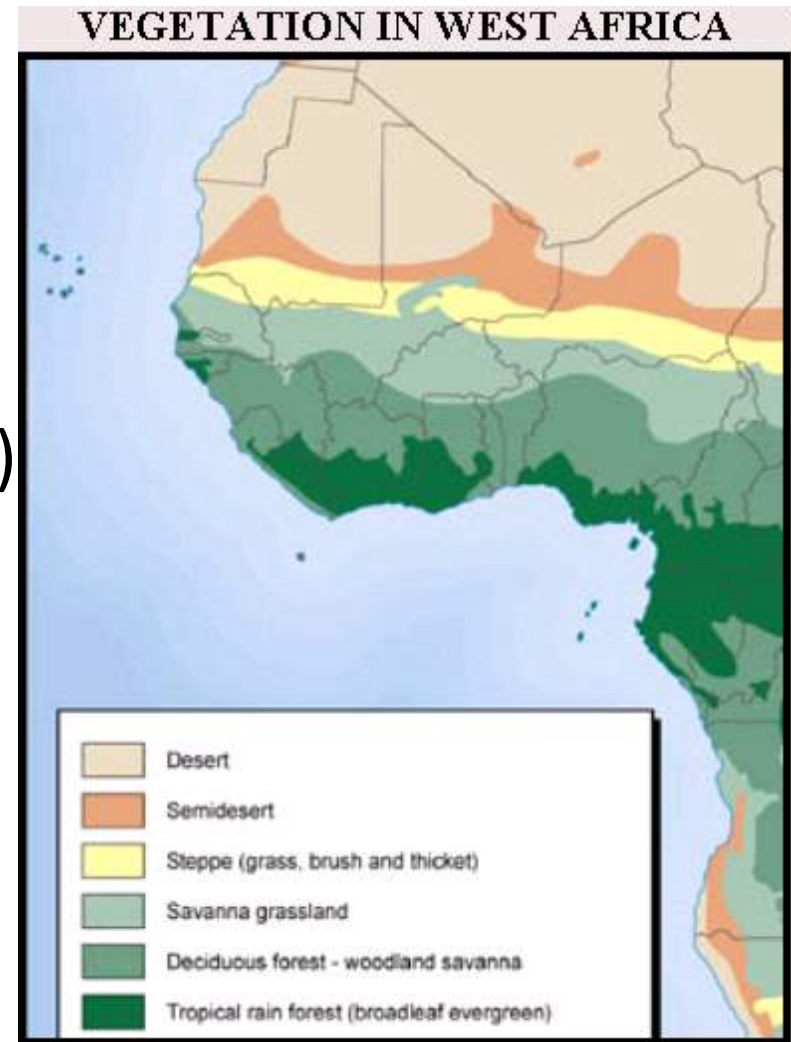
SASHA and the Sweetpotato Support Platform: Breeding and Seed Systems for West Africa

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6 July 2010

Sweetpotato Support Platform

West Africa

- Non-sweet Pre-breeding (unsweetpotato = USP)
 - Drawing on global capacity
 - NIRS
- Breeding Backstopping/
partnership (AGRA and others)
 - Ghana
 - Nigeria
 - Burkina Faso
 - Others
- Germplasm Exchange
Seed System Support



Even though it's not sweet the vehicle
has to be orange!





Virus resistant
Weevil resistant
Dual-purpose
OFSP

Kumasi

Kampala

Nairobi

Drought tolerant
OFSP

Mozambique

SPHI Central Base

Sweetpotato Support Platforms

Breeding Unsweetpotato

- Understanding USP
 - Sweetness
 - Population target (<12% total sugar, <9% sucrose)
 - Taste (sweetness, texture, appearance, etc)
 - Utilization – Participatory approach (producers, processors, consumers)
 - Adaptation (Major AEZs – Forest to Savanna)
 - Students – WACCI (Vivian Oduro; Ernest Baafi) and others

USP potential uncertain
Low dm absorbs too much oil



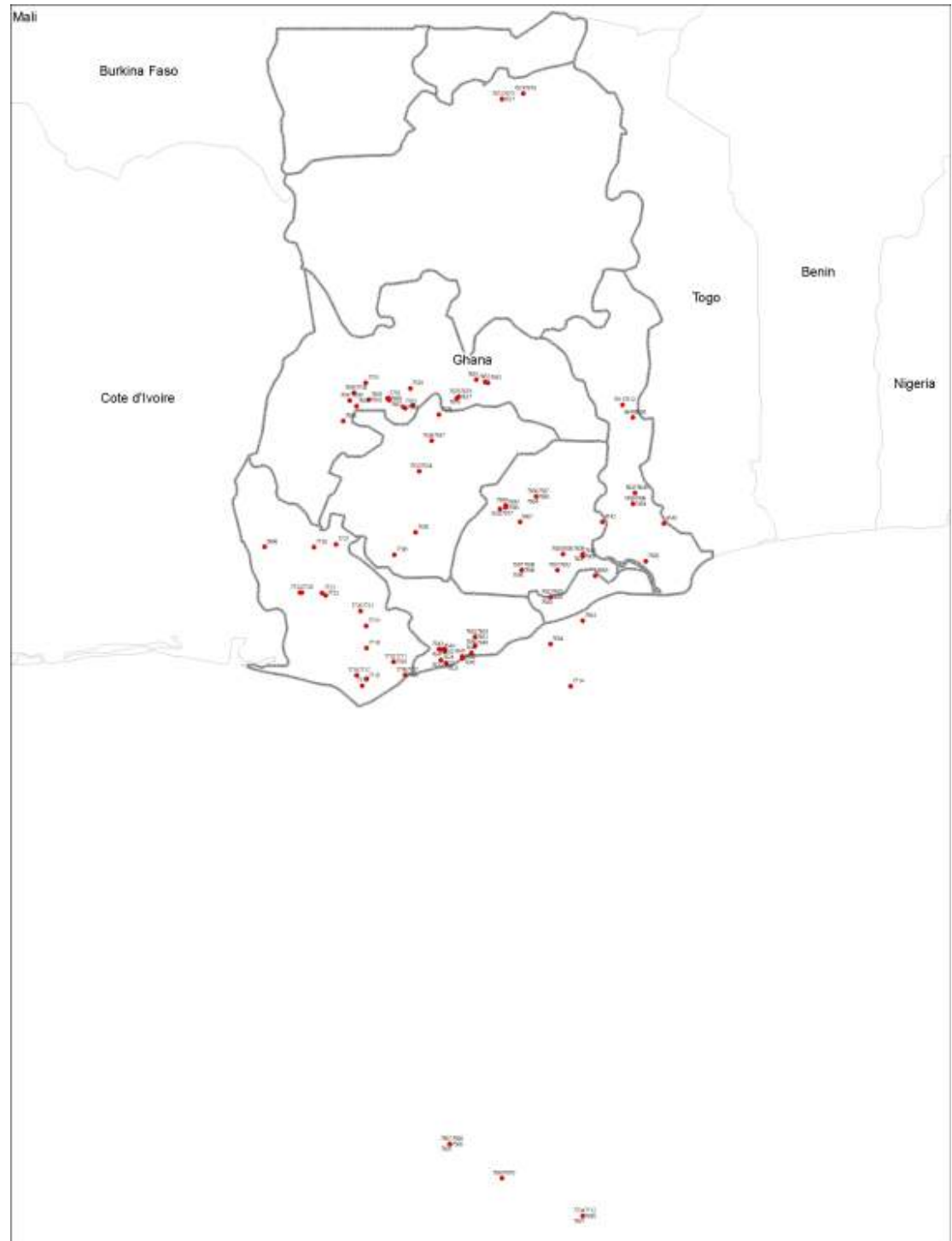
Understanding USP Germplasm sources

- West African Germplasm
 - Farmers varieties?
 - IITA materials
- Global genetic resources
 - CIP (Population B)
 - USDA (Stan Kays)
 - Kyukei 97 ~7% (bake) ~7% (raw), v. low amylase
 - CN1280-3 ~20% (bake) 9% (raw), v. low amylase

Germplasm

First map showing
distribution of
sweetpotato
germplasm
collection in
Ghana

Next steps are clear
Vivian Oduro



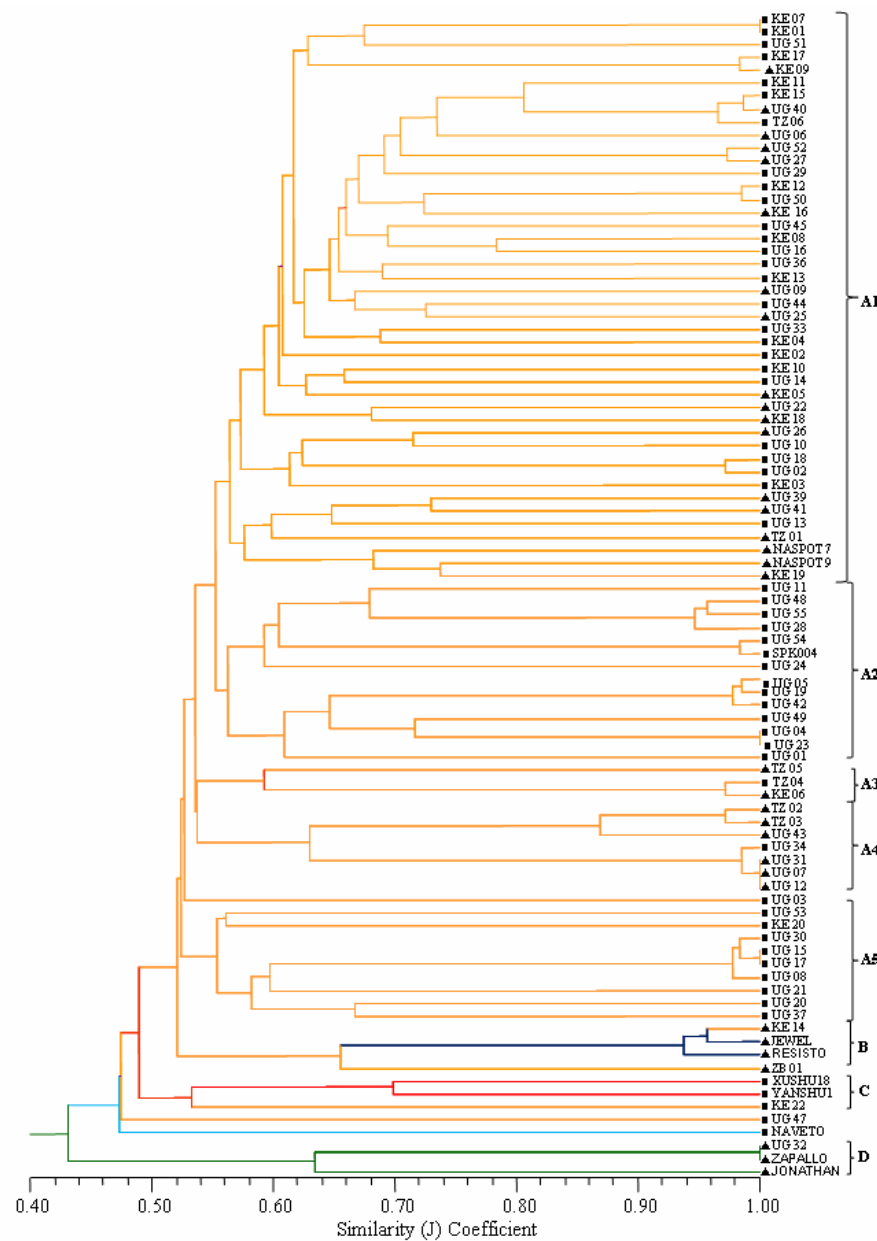
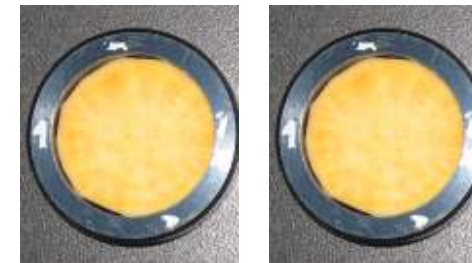
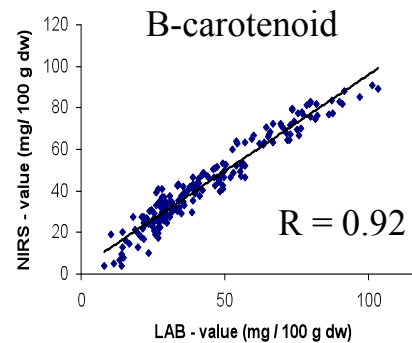
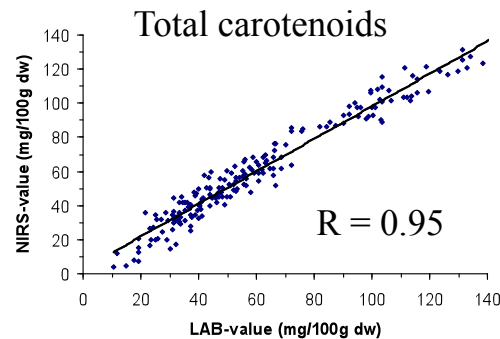


Fig. 2 Dendrogram of the UPGMA cluster analysis of 92 sweetpotato cultivars (85 cultivar of African origin) on the basis of Jaccard's SSR based genetic similarities

(origin of clones: orange = Africa, green lines = South American germplasm, light blue = Pacific, deep blue = North America, and red = China; origin of East Africa farmer varieties: UG = Ugandan, cultivars, KE = Kenya, TZ = Tanzania, ZB = Zambia; □ = White fleshed clones, Δ = Orange-fleshed clones)

(VII) NIRS (Near Infrared Reflectance Spectroscopy)



Sweetpotato dried roots:

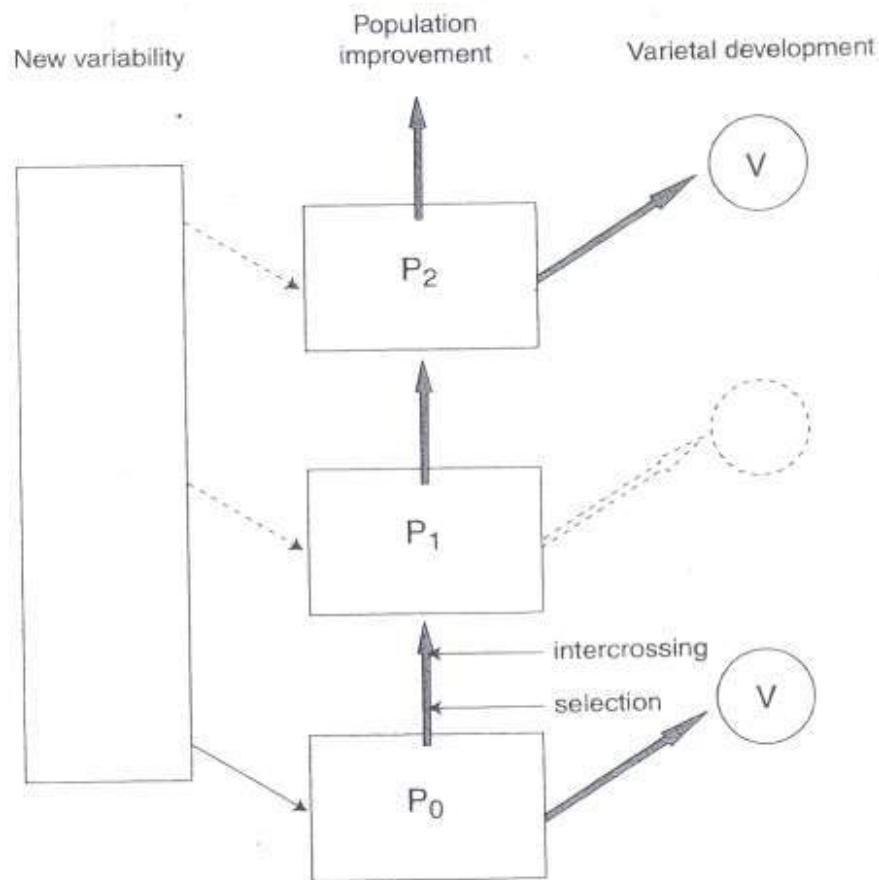
Protein, starch, sucrose, fructose, glucose, total carotenoids, b-carotene, Fe, Zn, Mg, Ca, Mg

Sweetpotato fresh roots:

Protein, starch, sucrose, fructose, glucose, total carotenoids, b-carotene, Fe, Zn, Mg, Ca, Mg

CIP is conducting the NIRS calibration development for Harvest Plus across all HP crops

Sweetpotato Support Platform (SSP) Breeding Activities



- Population (P) improvement by SSP – long term (SASHA)- use distinct populations to lead to major improvements (heterosis) in e.g. yield, disease resistance
- Variety (V) improvement by sweetpotato National programs – short term (AGRA or other support)

Seedling nursery and Crossing Block



Multiplication for trials with SARI



Kumasi is a virus hot spot

Apomuden - Resistant **Jewel - Susceptible**



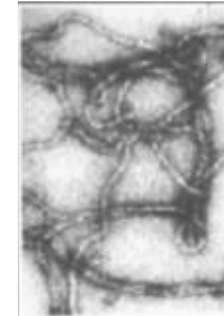
Markers and Sweetpotato virus disease (SPVD)



Aphids



SPFMV + SPCSV →
and / or Crinivirus
other
viruses



Whiteflies



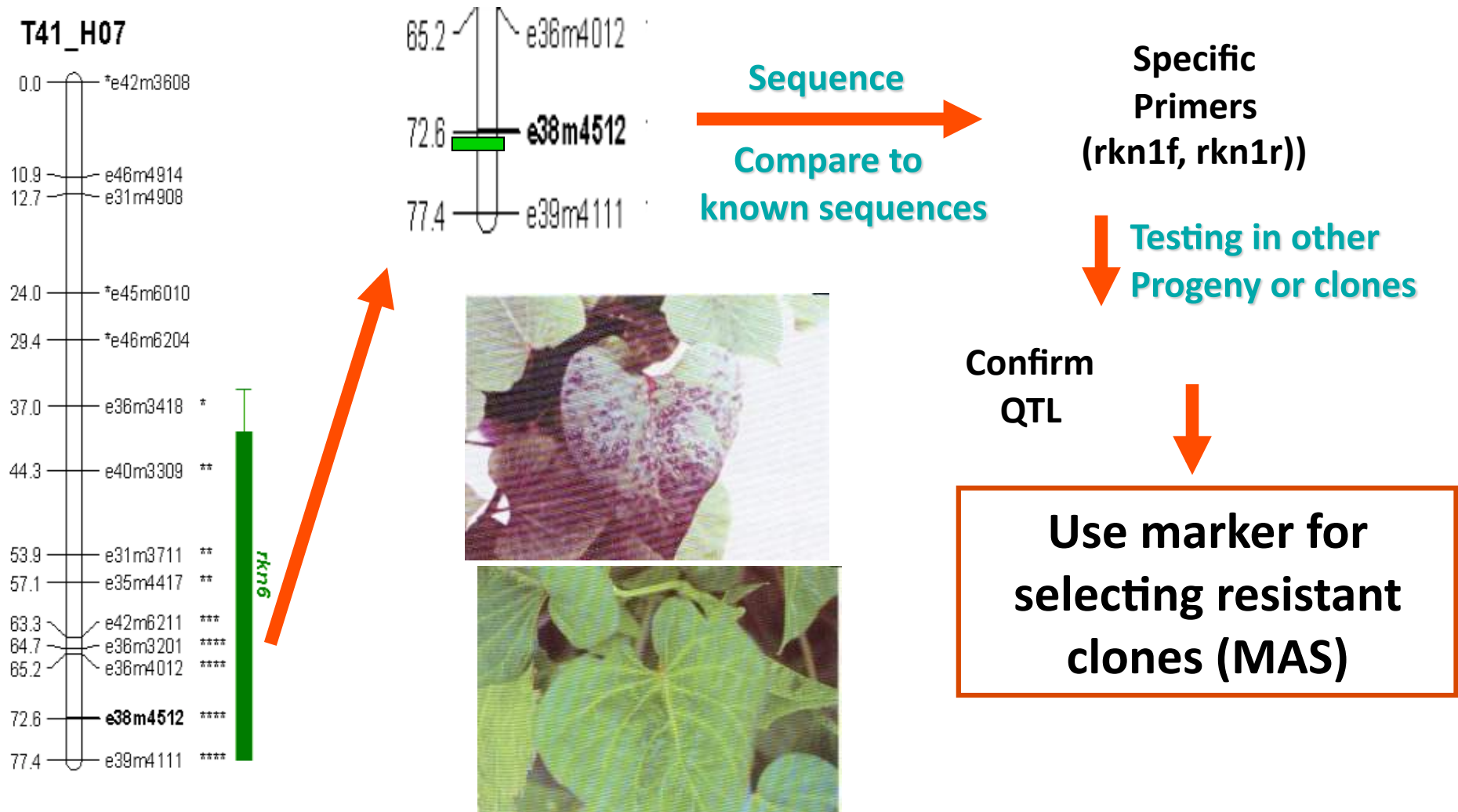
Apparently healthy
SPFMV and / or other
viruses

SPVD
SPCSV + SPFMV and / or
Other viruses

In experiments:
Yield reduction up to 90%

In practice:
Yield reduction up to 75%

Identify Gene/Marker and Verify Marker

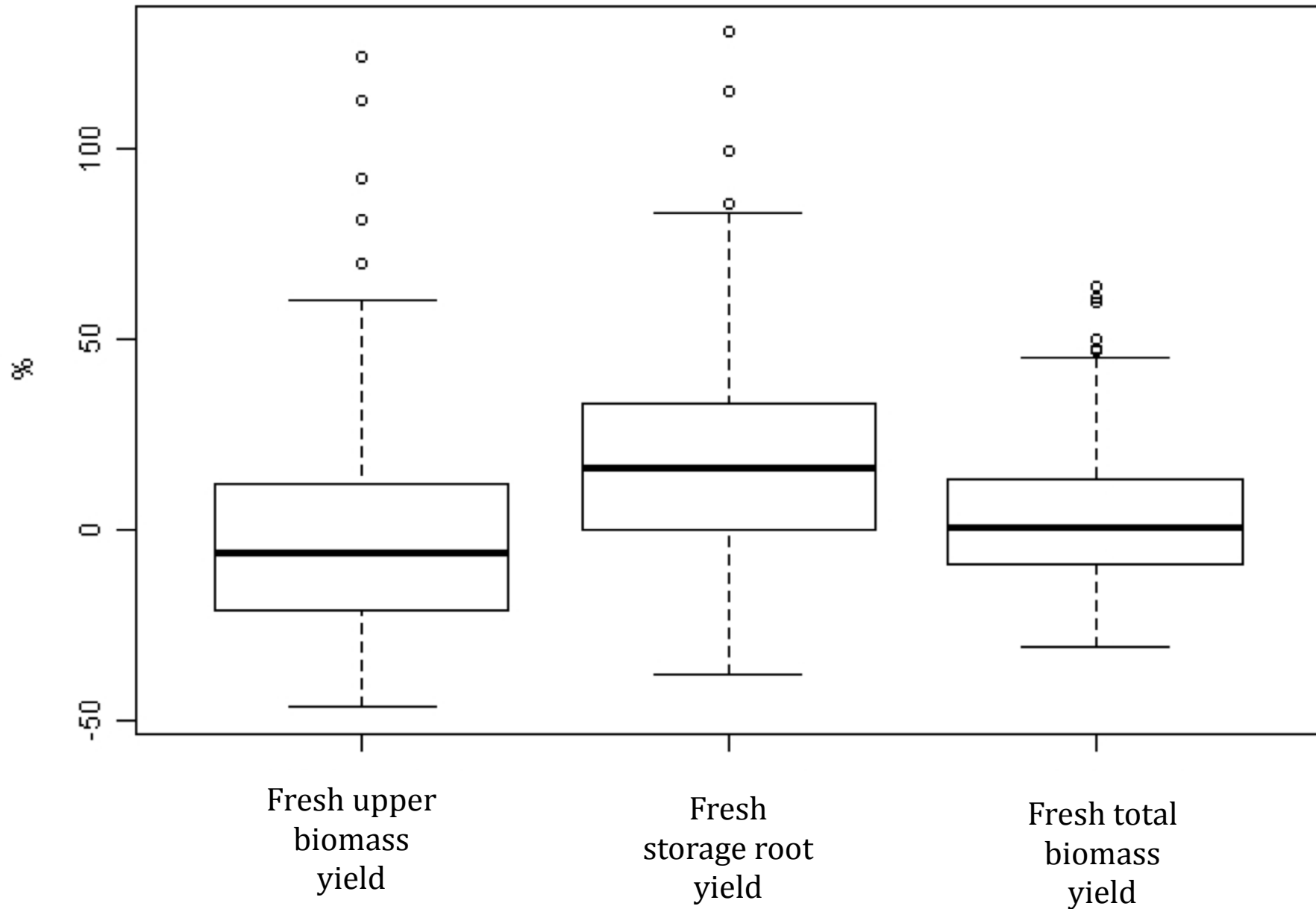


Planting early selection stages of sweetpotato for the accelerated breeding scheme in San Ramon (one of 3 locations)



Plot size: 1m row plot

Mid-parent – progeny distributions 231 families



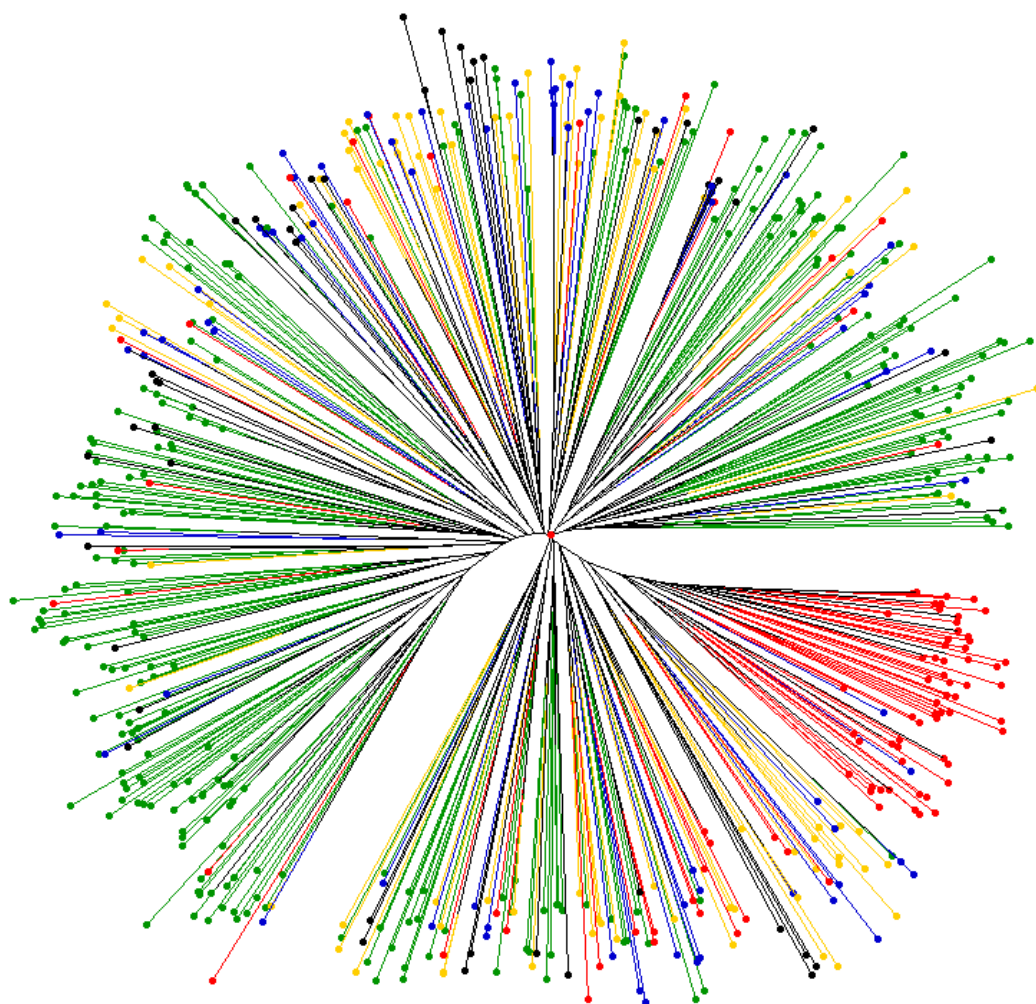
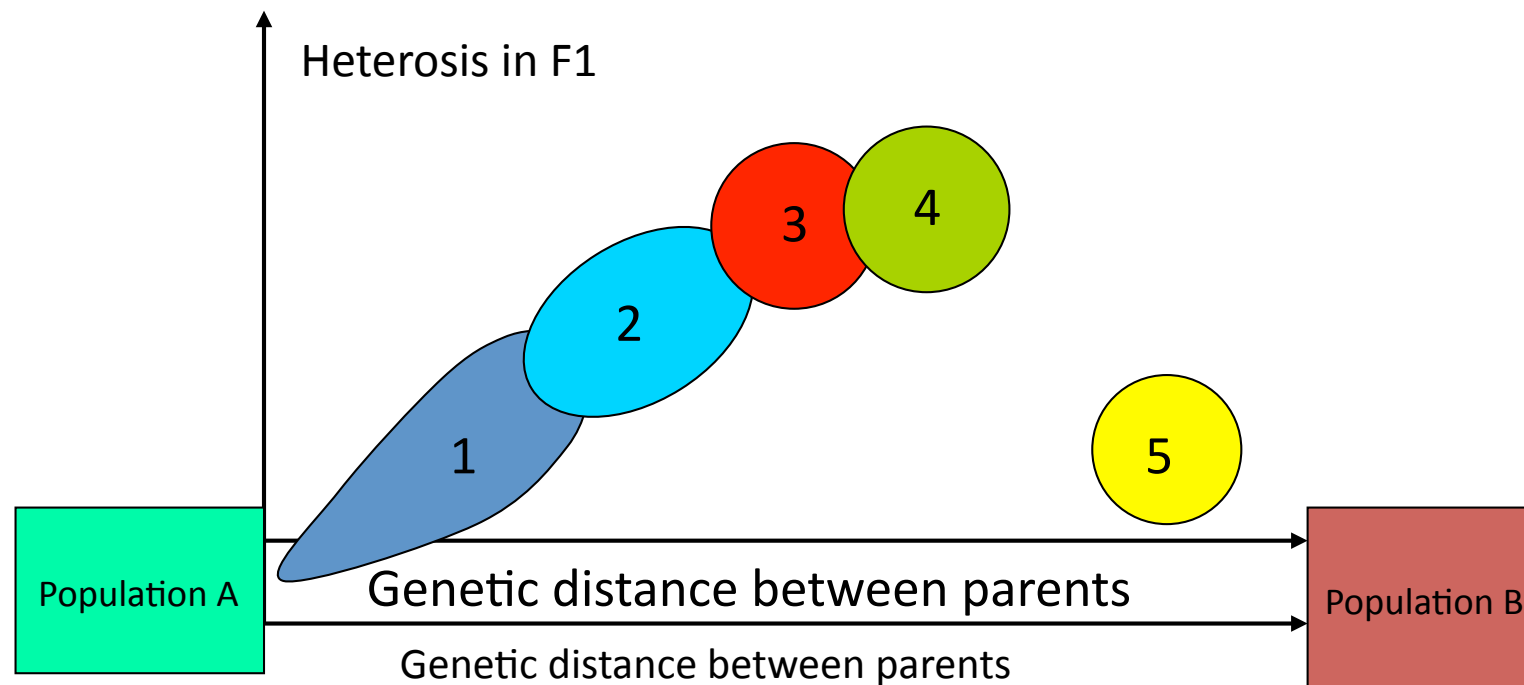


Fig. Cluster analysis of 575 accessions of sweetpotato based on 20 SSR markers

(Central and North America in black, South America in green, Pacific including New Zealand, Austria and Papua New Guinea in blue, Asia in yellow and Sub Saharan Africa in red).

Marker Application for Heterosis and Genetic Distance

Heterosis increases with the Genetic Distance
& decreases if the Genetic distance is too large



Schematic representation of the relationship between mid-parent heterosis for yield and parental genetic distance based on unselected DNA markers (1) Crosses between related parents (2) intra-genepool crosses between unrelated lines (3) - (5) inter-genepool crosses (Lefort-Buson 1985, Melchinger 1999, modified)

Germplasm Exchange + Seed System Support



Example: Recognized viruses that infect sweetpotato

| Genus | Virus | Transmission | Distribution |
|---------------------|--------|--------------|--|
| <i>Potyvirus</i> | SPFMV | Aphid | Worldwide |
| | SPLV | Aphid | Taiwan, China, Japan, Indonesia, Philippines, India, Egypt |
| | SPMSV | Aphid | Argentina, Peru, Indonesia, Philippines, China, Egypt, South Africa, Nigeria, New Zealand |
| | SPVG | Aphid | China, Japan, USA, Egypt, Ethiopia, Nigeria, Barbados, Peru, Spain, South Africa |
| | SPV2 | Aphid | USA, Taiwan, China, South Africa, Portugal, Australia, Barbados |
| | SPCSV? | Unknown | Caribbean Region, Zimbabwe, Uganda, Kenya |
| | SPVMV | Aphid | Argentina |
| <i>Ipomovirus</i> | SPMMV | Whitefly? | Africa, Indonesia, China, PNG, India, Egypt, New Zealand |
| | SPYDV | Whitefly | Taiwan, Far East |
| <i>Crinivirus</i> | SPCSV | Whitefly | Worldwide |
| <i>Cucumovirus</i> | CMV | Aphid | Israel, Egypt, Kenya, South Africa, Japan, New Zealand |
| <i>Begomovirus</i> | SPLCV | Whitefly | Far East, USA, China, Taiwan, Japan, Korea, Europe, Africa?, Peru |
| | SPLCGV | Whitefly | USA, Puerto Rico |
| | IYVV | Whitefly | Spain, Italy |
| | ICLCV | Whitefly | Israel |
| <i>Carlavirus</i> | SPCFV | Unknown | Africa, China, Taiwan, North Korea, Cuba, Panama, South Americaa, N. Zealand |
| | C-6? | Unknown | USA, Peru, Cuba, Dom. Rep., Indonesia, Philippines, P. Rico, Egypt, Kenya, South Africa, New Zealand |
| <i>Nepovirus</i> | SPRSV | Unknown | Papua New Guinea, Kenya? |
| <i>Caulimovirus</i> | SPCaLV | Unknown | South Pacific Region, Madeira, China, Egypt, P. Rico, Nigeria, Kenya? |
| <i>Ilarvirus</i> | TSV | Unknown | Guatemala |
| <i>Polerovirus</i> | SPLSV | Aphid | Peru, Cuba |
| <i>Tobamovirus</i> | TMV | None | USA |
| Unknown | C-3 | Unknown | Brazil. Unknown in others countries |
| | C-9 | Unknown | ? |

Courtesy: Segundo, CIP

Importance for seed systems

- Recognized potential impact on poverty
- Impact on yield/quality/income
 - Provision of clean planting material alone can yield rates of return of between 56-84%.
- Pathway for dissemination of new varieties
- Provision of **timely** (clean) planting material of appropriate varieties to extend availability of the crop and in times of crisis

The Sweetpotato Community of Practice survey questionnaire indicated that “**quality and availability of planting material**” is the most important limiting factor in developing the sweetpotato crop in all three sub-regions

Virus-free sweetpotato planting material in Shandong province, China

- CIP technology (virus testing and tissue culture) transferred to China (1988-1998)
- Technology adopted over 80% of area (17Mt; 12% global): 1994-1998. Average yield increase of 30%
- Annual productivity increases by 1998 valued at \$145M p.a. (NPV \$550M; IRR of 202%)
- Agricultural income of 7M smallholders improved by 3-4%.

Varieties vary in their ability to resist viruses over time



Viruses more of a problem in areas where sweetpotato is grown year-round and aphid and white fly pressure is high

Requirements for high quality foundation (pre-basic) planting usually maintained by national programs

- Critical enabling technologies.
 - In vitro conservation of germplasm
 - Virus clean up capacity (thermotherapy)
 - Virus diagnostic capacity (safe movement)
- In vitro multiplication capacity
- Screenhouse and field multiplication of clean vines



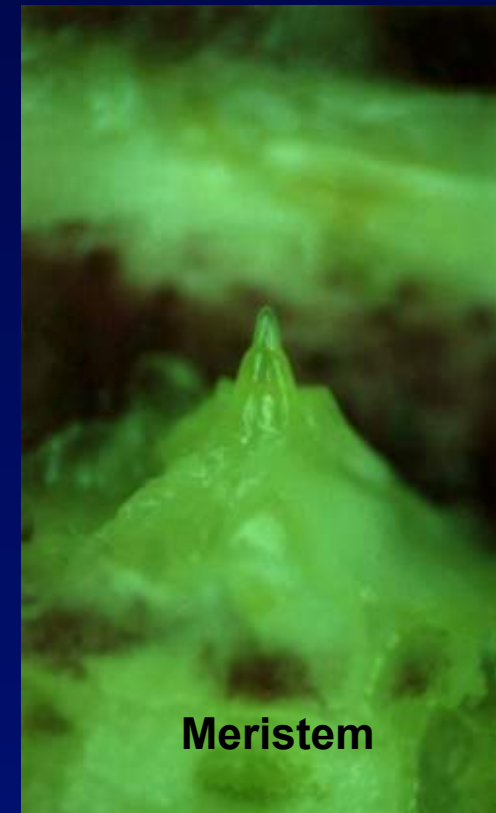
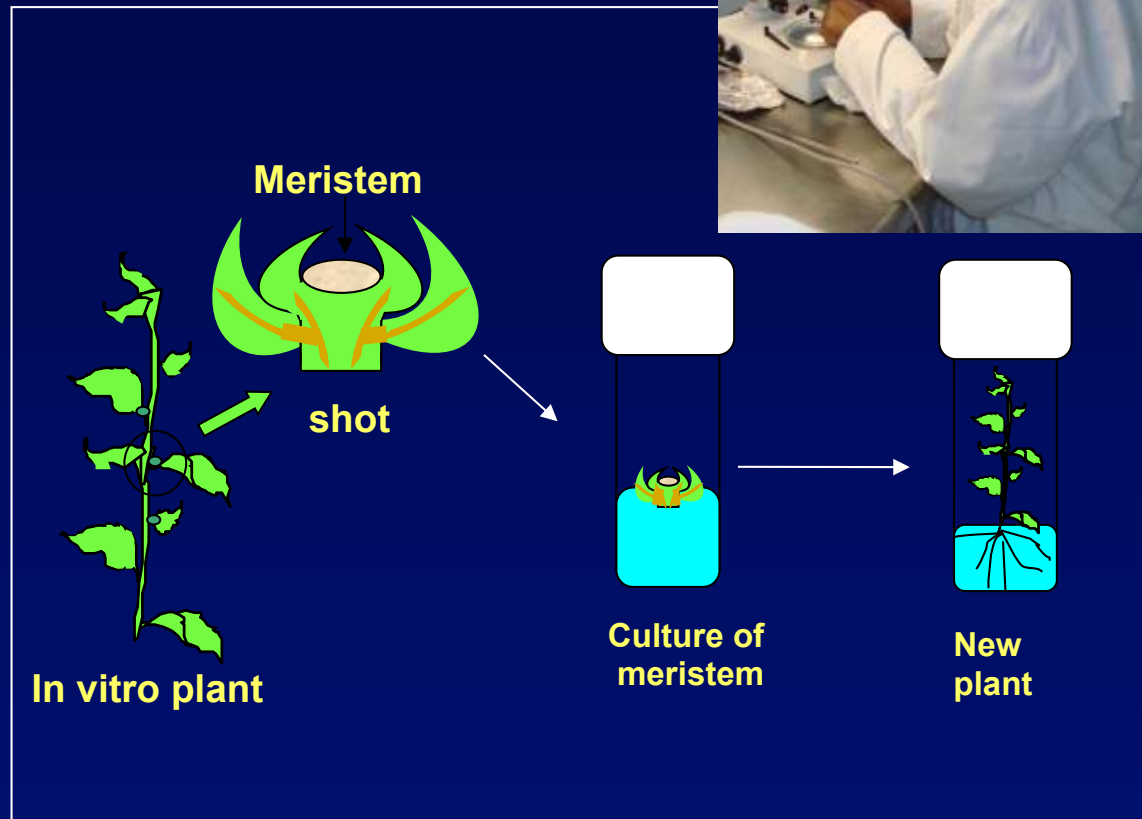
Thermotherapy (34-36°C for 1 month)



Growth chamber

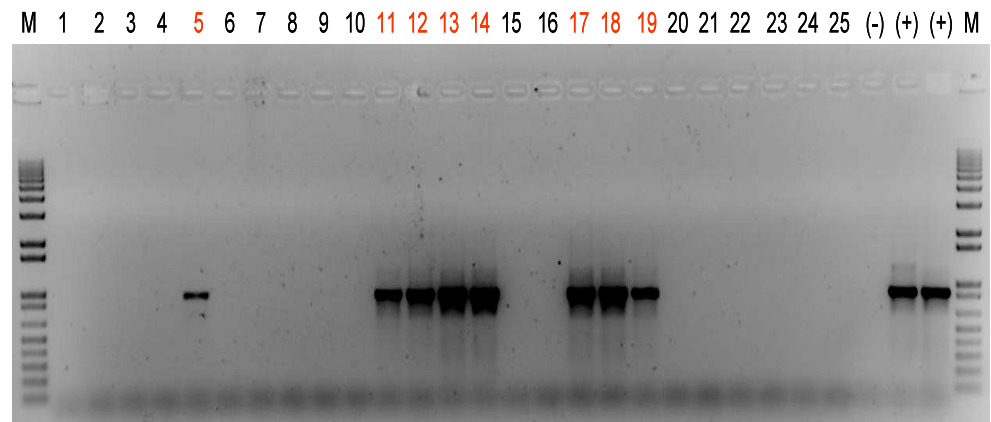


Meristem culture



Virus indexing

- Cleaned plant are first indexed for viruses through:
 - Grafting in *I. setosa*
 - NCM ELISA
 - PCR



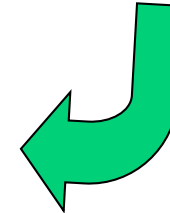
Diagnostic equipments



Realtime PCR



Growing plantlets in a greenhouse



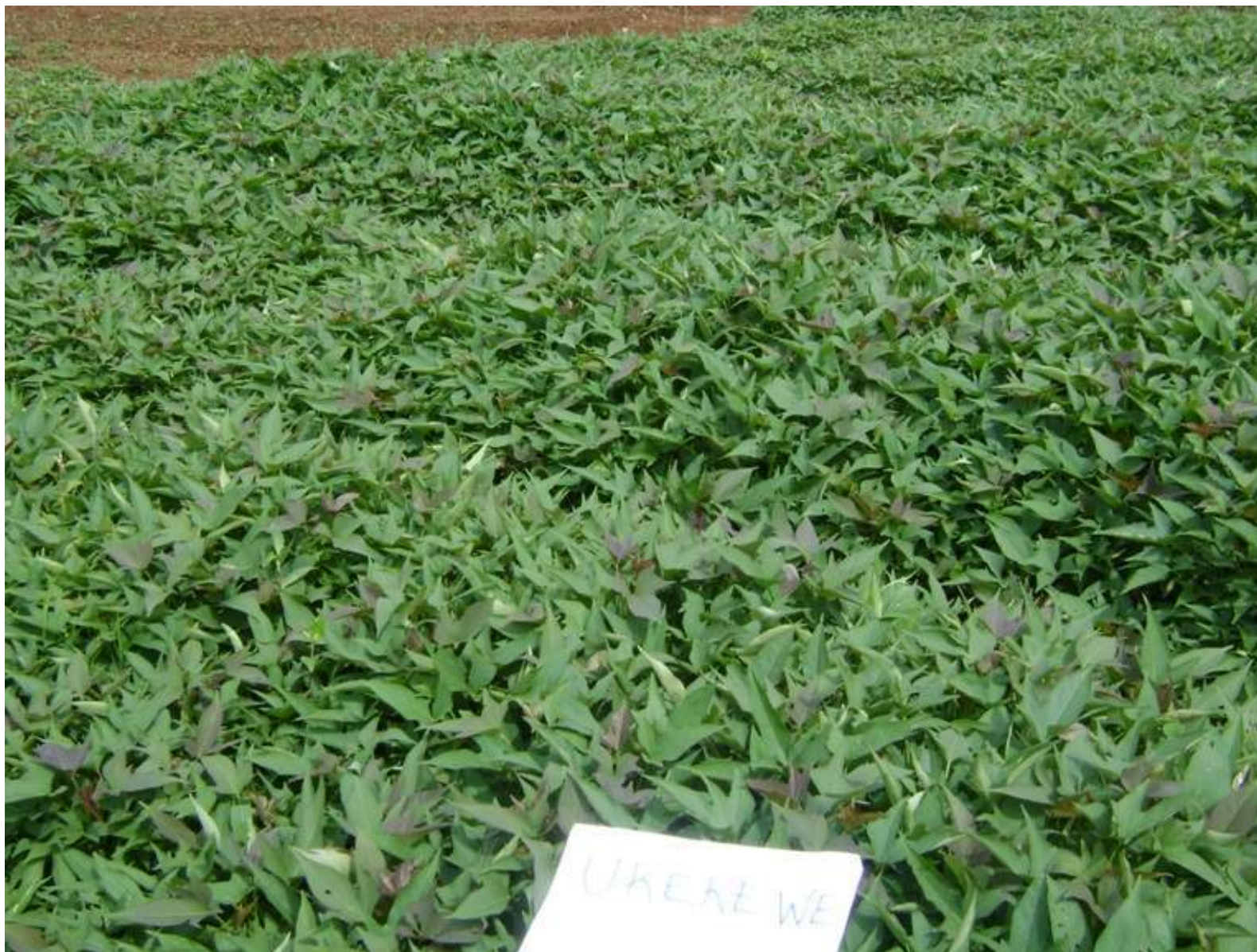
Plantlets for hardening



Transferred to Hardening shade



Ukerewe – Primary multiplication beds



Ejumula – Primary multiplication beds



Identified Challenges

1. How to maintain vines during the dry season, particularly when access to lowlands is limited?

2. How to assure that adequate quantities of quality vines are available at the beginning of the rains?

- Results from some experiments to date start to answer these questions

Strategies to conserve vines through dry seasons and extend availability of crop: buried roots and sources of water



Protected Root Bed Research in Uganda Built upon A Traditional Practice

Tradition in dry areas: leave a few unharvested roots in the ground and when the rains re-start the next season, they sprout

Drawbacks:

- Small amounts of material that need to be further multiplied so miss the productivity gain that comes with planting during the initial rains
- Or have small plots
- Material may not be healthy

Selected small but healthy roots and harvest time and store in sand during the dry season (Namanda PhD research)



Sand prevents weevil attack, better quality root storage than ash



Plant roots 10 cm deep, slanted, 20 cm (between rows) X 10 cm in protected area 6-10 weeks prior to expected start of rains



Initial water applications to be fortnightly to keep the roots moist, and then gradually be increased to weekly as the demand from the foliage increases. Two weeks prior to harvesting, reduce watering to harden off the foliage. Biggest problem: pests!

Which technology is best for helping smallholders maintain vines during the dry season?



Treadle pumps require 2-3 people to manage and hard to repair; hose easily damaged; higher output with manual irrigation. Small-scale drip irrigation (bucket +100 sq m) promising. Output 36% higher than manual irrigation in one trial. Farmers may diversify use.

What rapid multiplication techniques can be adopted by farmer multipliers?



Existing recommendation:

- Short 2-3 node cuttings
- 10 cm X 10 cm spacing
- Intensive management
 - Just not adopted over time
 - Used at stations with resources

Revised recommendation (Uganda):

- Apply farm yard manure/compost at a rate of 2.5 kg m⁻² or NPK (25-5-5) at the rate of 100g per m² within 10-15cm depth (*fertilization doubles yields*)
- 20 cms cuttings, 2/3 below surface
- 20 X 10 cm spacing

35-50
Vines per
root

Managing for dual purpose use: maximizing root and vine yields in Mozambique

Experimental design:

- Varied vine harvesting times: 60, 100, 150 days after planting (dap)
- Length of cutting: 15 vs 30 cm
- Planting density: 15 cm X 100 cm vs 30 cm X 100 cm
- Two varieties: Resisto vs MgCL01
- No fertilization

Results from 1st year:

- Best combination (total roots + vines):
 - vine harvesting 100 dap
 - 15 cm length cuttings
 - 15 cm X 100 cm planting density
- Worst combination: 60 dap; 15 cm cuttings, 15 cm X 100 cm spacing
 - Early vine harvesting negatively affects root production

35-50
Vines per
root

Building a Community of Practice

- Automated breeding data management
- Hands-on experience



Thank-you for your Attention
and Hospitality

